

Research article

Open Access

The hemolytic and cytolytic activities of *Serratia marcescens* phospholipase A (PhIA) depend on lysophospholipid production by PhIA

Ken Shimuta¹, Makoto Ohnishi*¹, Sunao Iyoda¹, Naomasa Gotoh²,
Nobuo Koizumi¹ and Haruo Watanabe¹

Address: ¹Department of Bacteriology I, National Institute of Infectious Diseases, Toyama 1-23-1, Shinjuku-ku, Tokyo 162-8640, Japan and ²Department of Microbiology and Infection Control Sciences, Kyoto Pharmaceutical University, Misasagi-Nakauchicho 5, Yamashinaku, Kyoto, 607-8414, Japan

Email: Ken Shimuta - shimuta@nih.go.jp; Makoto Ohnishi* - ohnishi7@nih.go.jp; Sunao Iyoda - siyoda@nih.go.jp; Naomasa Gotoh - ngotoh@mb.kyoto-phu.ac.jp; Nobuo Koizumi - nkoizumi@nih.go.jp; Haruo Watanabe - haruwata@nih.go.jp

* Corresponding author

Published: 16 December 2009

Received: 29 May 2009

BMC Microbiology 2009, 9:261 doi:10.1186/1471-2180-9-261

Accepted: 16 December 2009

This article is available from: <http://www.biomedcentral.com/1471-2180/9/261>

© 2009 Shimuta et al; licensee BioMed Central Ltd.

This is an Open Access article distributed under the terms of the Creative Commons Attribution License (<http://creativecommons.org/licenses/by/2.0>), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Abstract

Background: *Serratia marcescens* is a gram-negative bacterium and often causes nosocomial infections. There have been few studies of the virulence factors of this bacterium. The only *S. marcescens* hemolytic and cytotoxic factor reported, thus far, is the hemolysin ShIA.

Results: An *S. marcescens* *shlAB* deletion mutant was constructed and shown to have no contact hemolytic activity. However, the deletion mutant retained hemolytic activity on human blood agar plates, indicating the presence of another *S. marcescens* hemolytic factor. Functional cloning of *S. marcescens* identified a phospholipase A (PhIA) with hemolytic activity on human blood agar plates. A *phlAB* deletion mutant lost hemolytic activity on human blood agar plates. Purified recombinant PhIA hydrolyzed several types of phospholipids and exhibited phospholipase A1 (PLA1), but not phospholipase A2 (PLA2), activity. The cytotoxic and hemolytic activities of PhIA both required phospholipids as substrates.

Conclusion: We have shown that the *S. marcescens* *phlA* gene produces hemolysis on human blood agar plates. PhIA induces destabilization of target cell membranes in the presence of phospholipids. Our results indicated that the lysophospholipids produced by PhIA affected cell membranes resulting in hemolysis and cell death.

Background

Serratia marcescens is widely distributed in natural environments and has emerged in the last two decades as an important nosocomial pathogen, mainly in immunocompromised patients [1,2]. Although *S. marcescens* pathogenicity is poorly understood, its extracellular secreted enzymes, including several types of proteases, are candi-

dates for virulence factors [2]. Other factors (e.g., fimbria for adhesion, lipopolysaccharide (LPS), and ShIA hemolysin) have also been suggested as virulence factors [2,3].

Hemolysins are produced by various pathogenic bacteria and have been proposed to be responsible for their pathogenesis [4-6]. These hemolysins, including *S. marcescens*

ShlA, also have cytolytic activity [7]. One type of hemolysin/cytolysin is a group of pore-forming toxins. This type of toxin typically forms a homo-oligomer integrated into its target cell membrane, thereby changing the cell permeability and leading to cell death. ShlA has been shown to increase cell membrane permeability, but not to form an oligomer [3]. Another type of hemolysin has phospholipase C (PLC) activity. The α -toxin produced by *Clostridium perfringens* is the most thoroughly investigated PLC, but the molecular mechanism for its disruption of red blood cells (RBC) is not fully understood [8]. The pathogenic effects of other types of phospholipases, such as phospholipase A (PLA), have been studied in various bacteria, including *Helicobacter pylori* (PldA) [9], *Legionella pneumophila* (PlaA) [10], *Campylobacter coli* (PldA) [11], and *Yersinia enterocolitica* (YplA) [12]. Two extracellular PLAs, PhlA and PlaA, have been described previously in *Serratia* species [13,14]. PlaA is produced in *Serratia* sp. strain MK1 isolated from Korean soil [14]. The amino acid sequence of PlaA was found to have significant similarity (80%) to PhlA from *S. marcescens* MG1, which was originally classified as *S. liquefaciens* [13-15]. However, the cytotoxic and hemolytic activities of these enzymes have remained unclear, and the importance of PLA in bacterial virulence is not well understood.

S. marcescens produces two types of hemolysins: contact-dependent hemolysin and extracellular hemolysin [16]. The ShlA hemolysin has cytolytic and contact-dependent hemolytic activity, but little is known about the *S. marcescens* secreted hemolysin. The gene cassette responsible for the production and secretion of ShlA is *shlAB*, with *shlA* encoding the structural gene for hemolytic activity and *shlB* required for activation and secretion of ShlA in the presence of the cofactor phosphatidylethanolamine [17]. ShlA production is higher at 30°C than at 37°C [18].

In this study, we cloned an *S. marcescens* gene that produced hemolytic activity on human blood agar plates. The gene, designated *phlA*, encoded an extracellular PLA activity. We also showed that PhlA hydrolyzed phospholipid to lysophospholipid, which directly lysed human, horse, and sheep RBC and cultured cells.

Methods

Reagents

Taurocholic acid sodium salt hydrate and ethylenediamine-N,N'-diacetic acid (EDDA), L- α -phosphatidylcholine (PC) and L- α -phosphatidylethanolamine (PE) were purchased from Sigma. Cardiolipin (CL), L-3-phosphatidylinositol (PI) and sphingomyelin (SPM) and L- α -lysophosphatidylcholine (LPC) were purchased from Doosan Serdary Research Laboratories. Lecithin from egg yolk was purchased from Wako Chemicals. Phospholipase A2 derived from bovine pancreas was purchased from Sigma.

Bacterial strains, plasmids, and media

S. marcescens niid 298 strain is one of our reference strains for serotyping, which is originally isolated from urine. *E. coli* K-12 DH5 α and pBR322 were used for shotgun cloning. The pGEM-Easy vector (Promega) was used for cloning. pCold1 and pG-KJE8 (TaKaRa) were used as expression vectors in *E. coli* BL21 [F⁺, *ompT*, *hsdSB* (rB-mB⁻), *gal*, *dcm*] (TaKaRa). Unless otherwise specified, bacteria were grown in Luria broth (LB). Antibiotics were added as required for the following final concentrations: ampicillin, 200 μ g/ml; kanamycin, 100 μ g/ml; and chloramphenicol, 20 μ g/ml.

Peripheral human blood was obtained from healthy volunteers with their informed consent according to the guidelines laid down by the Ethics Committee of National Institute of Infectious Diseases. Horse and sheep blood were obtained from Kohjinbio. RBC were washed three times with phosphate-buffered saline (PBS) by centrifugation at 850 \times g for 10 min. Washed RBC were resuspended in PBS to a final concentration of 2.5% (vol/vol) and used for hemolytic activity assays. Blood agar plates contained 12.5 g Bacto-tryptone, 5 g NaCl, 5% (v/v) RBC, 10 mM CaCl₂, and 15 g agar (per liter), and the pH was adjusted to 7.2 [19]. PCY medium plates, which contained 10 g Bacto-tryptone, 5 g yeast extract, 5 g NaCl, 20 mM CaCl₂, 5 g taurocholic acid, 15 g egg yolk lecithin, and 15 g agar (per liter), were used for determining phospholipase activity [14].

Functional cloning

Shotgun cloning was used to identify hemolytic factors as follows. *S. marcescens* strain niid 298 genomic DNA was digested with *Sau*3AI, ligated into a pBR322 vector *Bam*HI site, and introduced into *E. coli* DH5 α . Colonies that induced hemolysis on human blood agar were selected as candidate hemolysin clones. The inserted fragments were amplified by PCR and complete fragment sequences were determined using a 3130 Genetic Analyzer (Applied Biosystems). The *S. marcescens* nucleotide sequences determined in this work have been deposited in the DDBJ/EMBL/GenBank databases under the following accession numbers: AB505202 and AB505203 for the *phlA* and *phlB* genes of *S. marcescens* niid 298.

Construction of mutant strains

A one-step inactivation method [19-21] was used to obtain *shlBA* and *phlAB* deletion mutants. For construction of a *shlBA* deletion mutant, PCR products were amplified from pKD4 [19] with primers ShlBA5' (5'-GGTTAACCTCATGGATTGGGCTGGCTGCCCCGGCG-GCCTTCATAGTGTAGGCTGGAGCTGCTTC-3') and ShlBA3' (5'-GCAAACTCCACGCCTGCCGTCATGCTTCATGCTACTGTCAGCAACATATGAATATCCTCCTTACT-3'), which flank the 5' and 3' termini of the *shlBA* gene with 45

bp homology, and electroporated into *S. marcescens* niid 298 carrying pKD46 [20]. For construction of a *phlAB* deletion mutant, primers PhlAB5' (5'-AGCGCCAGTAAGGCTATCGCCAGCGCCCGCCGCAAGCGACCCCTCATATGAATATCCTCCTTAGT-3') and PhlAB3' (5'-TGCCTAAGAAAAACCGCCTGTACAGGCGGTT TTTTATGGGCGTCATATGAATATCCTCCTTAGT-3') were used. The correct mutation was verified by PCR using three different primer sets as described previously [20].

Preparation of purified PhlA

The full-length *phlA* gene was obtained from *S. marcescens* niid 298 genomic DNA by PCR with primers phlA5' (5'-GAATTCATATGAGTATGCCTTAAGTTTACCTCTG-3') and phlA3' (5'-GCTATCTAGATCAGGCATTGGCCTTCGCCTC-3'). The 5'- and 3'-termini of the PCR product were *NdeI* and *XbaI* restriction enzyme sites, respectively. The PCR fragment cleaved by these restriction enzymes was inserted into *NdeI/XbaI*-digested pCold1, which has a histidine tag site at the 5'-terminus, and used to transform *E. coli* DH5 α . The resulting plasmid, pCold1-phlA, was introduced into *E. coli* BL21 harboring pG-KJE8 [22]. The transformant was used for expression of His-PhlA according to the manufacturer's instructions.

To purify the His-PhlA recombinant protein, cells were harvested, lysed by lysis buffer [50 mM NaH₂PO₄, 300 mM NaCl, 10 mM imidazole, and protease inhibitors (one tablet of inhibitor mixture (Complete, Roche)/25 ml)], incubated in a final concentration of 1 mg lysis-enzyme/ml for 20 min on ice, and then disrupted by sonication. Cell debris was removed by centrifugation. The supernatant was analyzed by affinity chromatography using Ni-NTA agarose (Qiagen) under native conditions without a protease inhibitor. After dialysis against PBS, the purified protein was concentrated by Amicon Ultra-15 (MWCO = 30 K; Millipore). The protein concentration was determined using a BCA Protein Assay Kit (Pierce). We obtained approximately 1.9 mg purified recombinant protein (His-PhlA) from one liter of culture. The purified protein was analyzed by sodium dodecyl sulfate-polyacrylamide gel electrophoresis (SDS-PAGE) with Coomassie blue staining.

Determination of hemolytic activity

The assay for contact-dependent hemolysis was performed as described previously [23]. Briefly, *S. marcescens* cells were cultured in LB containing EDDA (2 mM) at 30°C or 37°C and harvested at log phase. Bacteria (1.2 × 10⁸ cells in 50 μ l PBS) were mixed with 70 μ l RBC and centrifuged (500 × g for 1 min). The mixture was incubated for 60 min at 30°C or 37°C with shaking. Hemoglobin released from lysed RBC was measured spectrophotometrically at 405 nm. Osmotic lysis of RBC in distilled water was taken as 100% hemolysis.

The hemolytic activity of purified PhlA in solution was measured as described previously [24,25], with the following modification. The RBC suspension containing 0.15 mg lecithin/ml, 0.06% taurocholic acid and 2 mM CaCl₂ was incubated with His-PhlA at 37°C for 1 h. After centrifugation (500 × g for 10 min) the supernatant was assayed spectrophotometrically. RBC were not lysed by this low concentration of taurocholic acid.

Detection of phospholipase A activity

Fluorogenic, BODIPY FL-labeled, phospholipase A substrates bis-BODIPY FL C₁₁-PC, PED6, and PED-A1 (Invitrogen) were used to determine the specificities of PLA1 and PLA2. The bis-BODIPY FL C₁₁-PC is glycerophosphocholine with BODIPY FL dye-labeled sn-1 and sn-2 acyl chains. PED-A1 and PED6 are glycerophosphoethanolamine with dye-labeled sn-1 and sn-2 acyl chains, respectively. The bis-BODIPY FL C₁₁-PC was self-quenched, and PED-A1 and PED6 fluorescence was quenched by added dinitrophenol. Release of the fluorophores by acyl chain cleavage by either PLA1 or PLA2 results in increased fluorescence. Each substrate solution (45 nM) was prepared in 10 mM Tris-HCl (pH 8.0), 100 mM NaCl, and 10 mM CaCl₂ [26]. A 90 μ l sample of each substrate solution was incubated with various concentrations of enzymes (10 μ l) in 96-well plates for 6 min, and fluorescence intensity was measured. The fluorescence background for each quenched substrate solution was determined without PhlA treatment. Fluorescence intensity was measured at 485 nm excitation and 530 nm emission using an Appliskan fluorescence microplate reader (Thermo Electron Corporation).

Assay for free fatty acids from phospholipids

Non-esterified fatty acids (NEFA) released from phospholipids (PLs) were quantitated by an enzymatic colorimetric method using a NEFA-C kit (Wako chemical, Japan) [27]. Substrate solutions were prepared by dissolving 5 mg of various phospholipids in 1 ml of a solution of 2% taurocholic acid and 10 mM CaCl₂. A 29 μ l sample of each substrate solution was mixed with 1 μ l His-PhlA and incubated at 37°C for 1 h. Background NEFA absorbance was estimated using non-His-PhlA treated substrates. NEFA concentrations were calculated from a calibration curve determined using oleic acid as a standard.

Thin-layer chromatography

PC (0.65 mM) was incubated with 8.3 μ M His-PhlA at 37°C for 1 h in the presence of 2% taurocholic acid and 10 mM CaCl₂. The reaction was terminated by placing the samples on ice. The samples were dried with a SpeedVac concentrator, dissolved in 20 μ l chloroform/methanol (2/1, vol/vol), applied to thin-layer chromatography plates (PE SILG; Whatman), and separated with chloroform/methanol/water (65/25/4, vol/vol/vol). LPC (1.0 mM)

was analyzed on the same plate as a reference. Phospholipids on the plate were visualized with Dittmer-Lester reagent [28].

Cell culture and cytotoxicity

HeLa and 5637 cells (derived from a human cervical cancer and bladder carcinoma, respectively) were grown in Dulbecco's Modified Eagle's Medium (DMEM) and 1640 RPMI medium, respectively, plus fetal calf serum (10% v/v) at 37°C in the presence of 5% CO₂. At 24 h before the start of cytotoxicity experiments, 96-well culture plates were seeded with 1.0×10^4 cells per well. After washing with medium, the cells were incubated with various concentrations of His-PhlA in 100 µl lecithin solution (313 µg/ml lecithin, 0.125% taurocholic acid, and 2 mM CaCl₂ in DMEM) at 37°C for 1 h. Cytotoxicity was measured as the amount of lactate dehydrogenase (LDH) released as determined with a CytoTox 96 Non-Radioactive Cytotoxicity Assay Kit (Promega) [29]. Complete (100%) cytotoxicity was determined by measuring LDH release after cell lysis with 1% Triton X-100.

Results

Identification of an *S. marcescens* hemolysin other than ShlA

S. marcescens niid 298 showed hemolytic activity visible as clear zones on human, sheep, and horse blood agar plates (Fig. 1A). The zones were larger for bacteria grown at 30°C than at 37°C. *S. marcescens* also showed contact-dependent hemolytic activity on human RBC, which was also greater for bacteria grown at 30°C than at 37°C (Fig. 1B).

Since ShlA is the only hemolysin that has been reported in *S. marcescens* [7], we constructed an *shlBA* deletion mutant. The mutant grown at both 30°C and 37°C lost its contact-dependent hemolytic activity (Fig. 1B), but retained hemolytic activity on human blood agar plates (Fig. 1C). These results indicated that *S. marcescens* had a hemolysin other than ShlA.

Functional cloning of a novel hemolysin

To clone the *S. marcescens* hemolysin identified on human blood agar, we constructed a library of *S. marcescens* strain niid 298 DNA in *E. coli* DH5α. Approximately 13,000 transformants were examined for hemolytic activity on human blood agar plates and four hemolysin-positive clones were obtained. One clone showed hemolytic activity on human, sheep, and horse blood agar plates, but the other three clones showed activity only on human blood agar.

Sequence analysis of the inserts in the three clones with hemolytic activity only on human blood agar showed that all three had *phlA* and *phlB* genes with nucleotide similar-

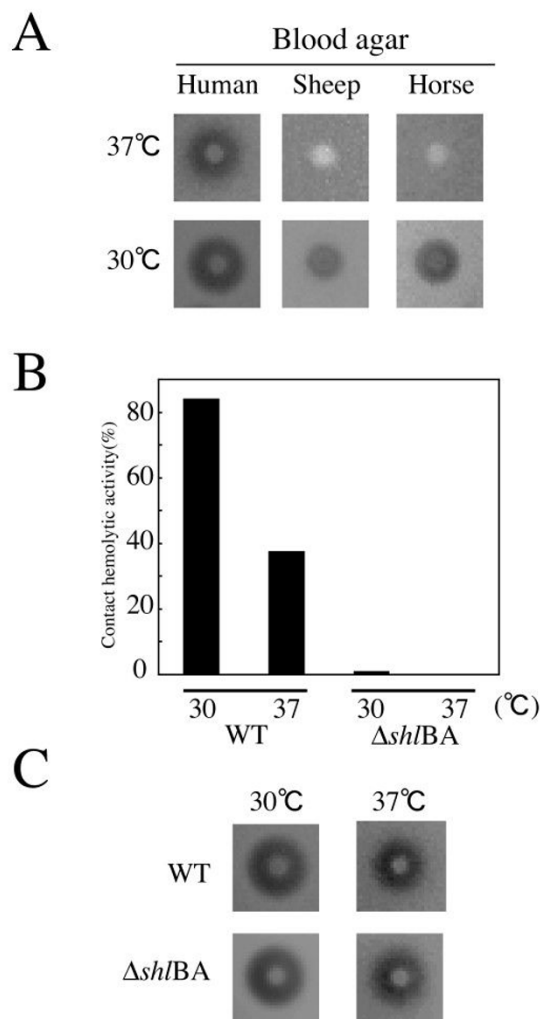


Figure 1
Hemolytic activity of *S. marcescens*. (A) Hemolytic activity of *S. marcescens* strain niid 298 on several blood agars. Cells (1×10^6) were cultured overnight, and then inoculated on various blood agars and incubated at 30°C or 37°C for 16 h. Clear zones indicated hemolysis. (B) Contact hemolysis assay for human RBC. Cells harvested in log phase were mixed with washed human RBC and incubated at 30°C or 37°C for 1 h with shaking. Released hemoglobin was measured spectrophotometrically as absorbance at 405 nm. Results are shown as percent lysis compared to complete lysis of RBC in distilled water. (C) Hemolytic activity of the *shlBA* deletion mutant on human blood agar. Experiments were performed as in (A).

ity to *phlA* and *phlB* (94% and 94%, respectively) of *S. marcescens* MG1, which was originally classified as *S. liquefaciens* [13,15]. The *phlA* and *phlB* deduced amino acid sequences were similar to *Serratia* sp. MK1 PlaA and PlaB (81% and 73% identity) and *Y. enterocolitica* Yp1A and Yp1B (60% and 50% identity) [12,14]. PhlB has been sug-

gested to be an inhibitor of PhIA inside the cell in which they are produced, thereby functioning to prevent PhIA activity until its release into the extracellular milieu [30]. Although there are no data about a PhIA hemolytic activity, since some other phospholipases have hemolytic activity, we investigated whether the *S. marcescens* *phlA* gene product might be a hemolysin.

Hemolytic activity of *S. marcescens* PhIAB is on human blood agar

To confirm that *phlAB* had phospholipase and hemolytic activities, we constructed the *phlAB* expression vector pGEMeas-y-*phlAB* and introduced it into *E. coli* DH5 α . *E. coli* DH5 α /pGEMeas-y-*phlAB* showed a clear zone on PCY agar plates containing egg yolk lecithin as a substrate for phospholipase, in contrast to *E. coli* DH5 α carrying an empty vector, indicating that PhIAB produced in *E. coli* DH5 α /pGEMeas-y-*phlAB* degraded phospholipids (Fig. 2A). In addition to phospholipase activity, *E. coli* DH5 α /pGEMeas-y-*phlAB* showed hemolytic activity on human blood agar plates (Fig. 2A).

We next constructed an *S. marcescens* niid 298 *phlAB* deletion mutant. The *S. marcescens* Δ *phlAB* mutant did not exhibit hemolytic activity on human blood agar plates or phospholipase activity on PCY agar plates (Fig. 2A). In contrast, the Δ *phlAB* mutant retained contact-dependent hemolytic activity against human RBC (data not shown). An *S. marcescens* Δ *phlAB* mutant carrying *phlAB* regained hemolytic and phospholipase activities (Fig. 2A), confirming that PhIAB had both activities.

Characterization of recombinant His-PhIA protein

To investigate PhIA hemolytic and phospholipase activities, we purified a recombinant His-PhIA protein produced in *E. coli* (Fig. 2B). Purified His-PhIA had hemolytic activity on human blood agar plates, but not on horse or sheep blood agar plates, and phospholipase activity on PCY agar plates (data not shown). These data indicated that PhIA had hemolytic and phospholipase activities, indicating that PhIB was not required for the PhIA activities.

We next studied the specificity of PhIA phospholipase. Phospholipase A (PLA) hydrolyzes the fatty acids of PLs at position *sn*-1 for phospholipase A1 (PLA1) and *sn*-2 for phospholipase A2 (PLA2), resulting in the release of free fatty acids and production of lysophospholipid (LPL). We measured free fatty acids after incubation of PhIA with various PLs [phosphatidylcholine (PC), cardiolipin (CL), L-3-phosphatidylinositol (PI), L- α -phosphatidylethanolamine (PE), and sphingomyelin (SPM)]. These experiments showed that PhIA cleaved ester bonds within PC, CL, PI, and PE and released fatty acids in a concentration-dependent manner, but did not hydrolyze SPM in our experimental conditions (Fig. 2C).

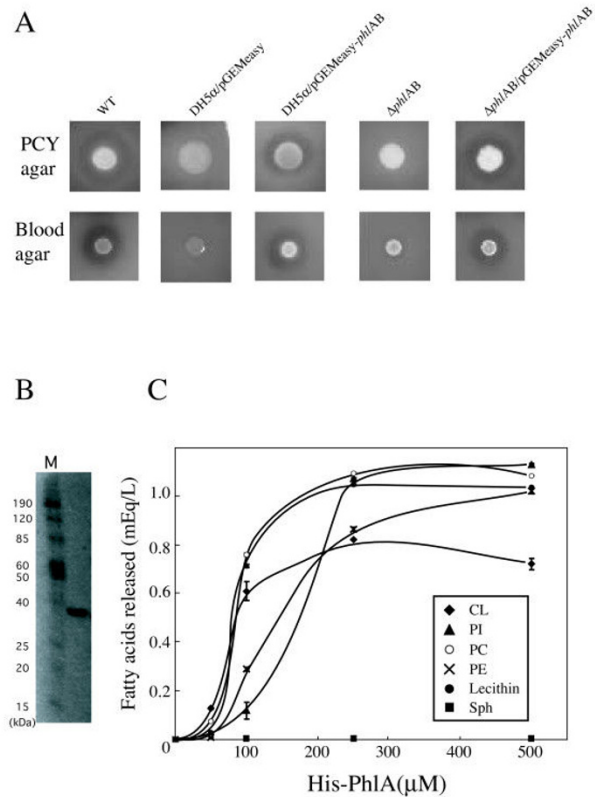


Figure 2
Phospholipase and hemolytic activities of *S. marcescens* PhIA. (A) Overnight cultures of wild-type strain *S. marcescens* niid 298, *E. coli* DH5 α cells carrying pGEMeas-y, *E. coli* DH5 α carrying pGEMeas-y-*phlAB*, *S. marcescens* niid 298 *phlAB* deletion mutant, and *S. marcescens* niid 298 *phlAB* deletion mutant carrying pGEMeas-y-*phlAB* (1×10^6 cells) were inoculated on blood agar plates and PCY agar plates and incubated at 37°C for 16 and 24 h, respectively. (B) Purified His-PhIA (1 μ g) was separated by 12.5% SDS-PAGE, and then was stained with Coomassie blue. Protein standards were in lane M, with relative molecular masses (kDa) at the left. (C) Various phospholipids were mixed with His-PhIA and incubated at 37°C for 1 h. Free fatty acids (FFA) released from phospholipids were detected using a NEFA-C kit. The amount of FFA was determined from an oleic acid calibration curve. Values are averages \pm SE of three independent experiments.

Previous reports have shown that some bacterial PLA2 enzymes have hemolytic activity [5,6,31]. However, there is little information on hemolysis caused by bacterial PLA1 enzymes. To confirm that *S. marcescens* PhIA had PLA1 activity, we tried to identify the site that is hydrolyzed by PhIA using fluorescent PLs as substrates [31,32]. As shown in Figure 3A, *S. marcescens* PhIA and bovine pancreatic PLA2 released fluorescent fatty acids from bis-BODIPY FLC11-PC, indicating that PhIA had phospholipase A activity (Fig. 3A). PhIA released fluorescent fatty

acids from PED-A1 in a concentration-dependent manner whereas control PLA2 did not produce fluorescence (Fig. 3B), indicating that PhIA was able to cleave ester bonds at PL sn-1 sites. Using PED-6 as substrate, although fluorescence intensity increased after PhIA treatment, the maximum fluorescence was 6-fold lower than after PLA2 treatment (Fig. 3C). These results are in agreement with the proposal that His-PhIA has PLA1, but not PLA2, activity.

PhIA requires phospholipids to express hemolytic activity

The studies described above showed that PhIA had hemolytic and phospholipase A1 activities. To investigate PhIA activity on a range of target cells, we studied the activity of purified PhIA in a solution reaction system with different types of cells. Interestingly, in contrast to the results on blood agar plates, PhIA hemolytic activity on human RBC was not detected in solution reactions, even at a PhIA concentration as high as 18 mM (Fig. 4A). This result indicated that PhIA did not act directly as a hemolysin on RBC. It has been reported that several animal venoms containing PLA exhibit an indirect hemolytic activity in the presence of lecithin [23,24]. When egg yolk lecithin or PC was added to the PhIA solution reaction system, PhIA was observed to have indirect hemolytic activity on human RBC (Fig. 4A).

Lysophospholipid (LPL) is one of the products from PLs hydrolyzed by PLA1. Therefore, we investigated whether LPL could cause hemolysis of human RBC. Lysophosphatidylcholine (LPC) was found to have hemolytic activity on human RBC in the solution reaction system (Fig. 4B). Using thin-layer chromatography, LPC was found to be produced by incubation of PC with PhIA (Fig. 4C).

To determine the range of cells affected by PhIA, we examined various kinds of RBCs. As described above, PhIA lysed human RBC, but not horse or sheep RBC, on blood agar plates. However, all three types of RBC were lysed by PhIA in a lecithin-dependent manner in the solution reaction system (Fig. 4D). An explanation of these results may be that, in human blood agar plates, enough PL might be released from collapsed RBC during agar plate preparation to allow PhIA to produce LPL. When PL was added to sheep and horse blood agar plates, we clearly detected PhIA-dependent hemolytic activity on horse and sheep blood agar plates (data not shown).

Since PC required 6-fold more PhIA than lecithin for induction of 50% hemolysis (Fig. 4A), the egg yolk lecithin used in this study might have contained enough LPL for hemolysis. However, no hemolysis was induced by lecithin without PhIA treatment (Fig. 4D).

Taken together, these results indicated that PhIA phospholipase activity hydrolyzed PL and produced LPL. Since

LPL is known to be a surfactant [33], it may have been the final effector leading to destabilization of the RBC membrane and hemolysis.

Cytotoxicity of PhIA in the presence of phospholipid

We examined the cytotoxicity of PhIA using HeLa and 5637 cells. PhIA had cytotoxic activity against both HeLa and 5637 cells in the presence of lecithin (Fig. 4E). To investigate the cytolytic activity of late log phase *S. marcescens* culture supernatants, *S. marcescens* was grown at 37°C for 6 h in LB containing PL. Up to 48-fold dilutions of the *S. marcescens* culture supernatant induced cell death of both HeLa and 5637 cells, while supernatant of *S. marcescens* Δ phlAB cultured under the same conditions had no effect on HeLa or 5637 cells, indicating that PhIA was an extracellular secretion product (data not shown).

Discussion

A wide range of pathogenic bacteria produce phospholipases, and the putative role of PLA in virulence has been studied in some of these pathogens. Outer membrane-associated PLAs (OMPLAs) were first identified in *E. coli* [34] and orthologs were subsequently reported in numerous gram-negative bacteria, including *H. pylori* (PldA) [9]. The OMPLAs have been well-characterized and are thought to enhance bacterial growth, colonization, and survival. In addition to modulation of the bacterial membrane, some OMPLAs were shown to have contact-dependent hemolytic/cytolytic activities [35]. Another group of PLAs (e.g., YplA [12], ExoU [36], PlaA [10], and SlaA [37]) is secreted from bacterial cells. Purified ExoU and SlaA [38,39] recombinant proteins do not show cytotoxic activity when added exogenously, and there is little information on the cytotoxicity of other secretory PLAs.

To our knowledge, ShIA is the only previously reported hemolysin from *S. marcescens*. Although, a Δ shlAB mutant showed hemolytic activity on blood agar plates, it did not exhibit contact-dependent hemolytic activity (Fig. 1C). Therefore, we performed functional cloning, which identified PhIA as an *S. marcescens* candidate hemolytic factor (Fig. 2A).

In the experiments reported here, we described the hemolytic and cytotoxic activities of *S. marcescens* PhIA. PhIA itself did not directly induce the destabilization of target cell membranes, but the LPL produced from PL by PhIA phospholipase activity showed hemolytic and cytolytic activities. Therefore, PhIA and ShIA have different hemolytic mechanisms. In addition, ShIA was expressed at lower temperature, but its expression decreased at 37°C [17]. In contrast, PhIA was expressed at 37°C, although its temperature regulation has not been elaborated. Also, PhIA hydrolyzed phosphoethanolamine (Fig. 3C), which is required for ShIA activity [16], implying that PhIA production could potentially regulate ShIA activity.

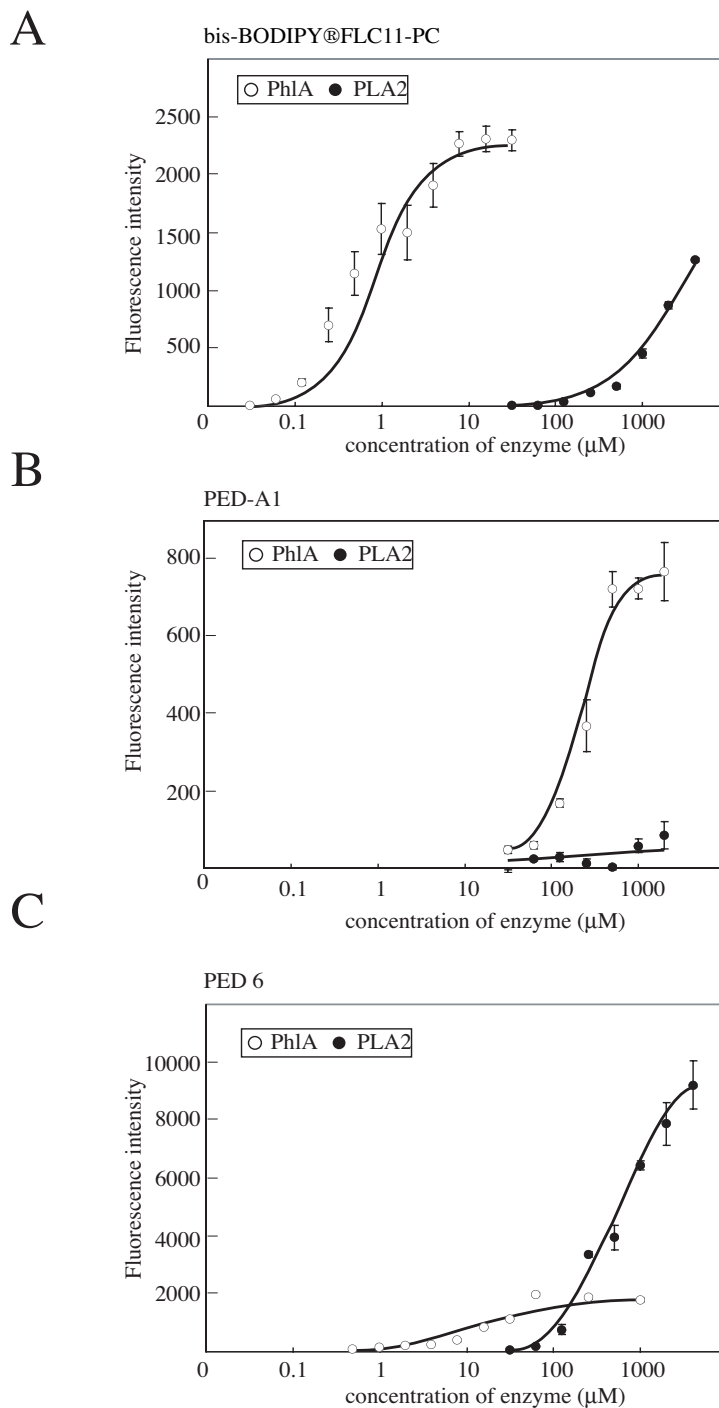


Figure 3
PLA1 and PLA2 activities of PhIA. PhIA activity was evaluated in a fluorescence enhancement assay using the following PLA fluorescence substrates: (A) bis-BODIPYFLC11-PC, (B) PED-A1, and (C) PED6. Fluorescence intensity was measured at 485 nm excitation and 530 nm emission using a fluorescence microplate reader (Appliskan; Thermo Electron Corporation). Open circles show His-PhIA; filled circles show PLA2 from bovine pancreas as a control. Values are averages ± SE from three independent experiments.

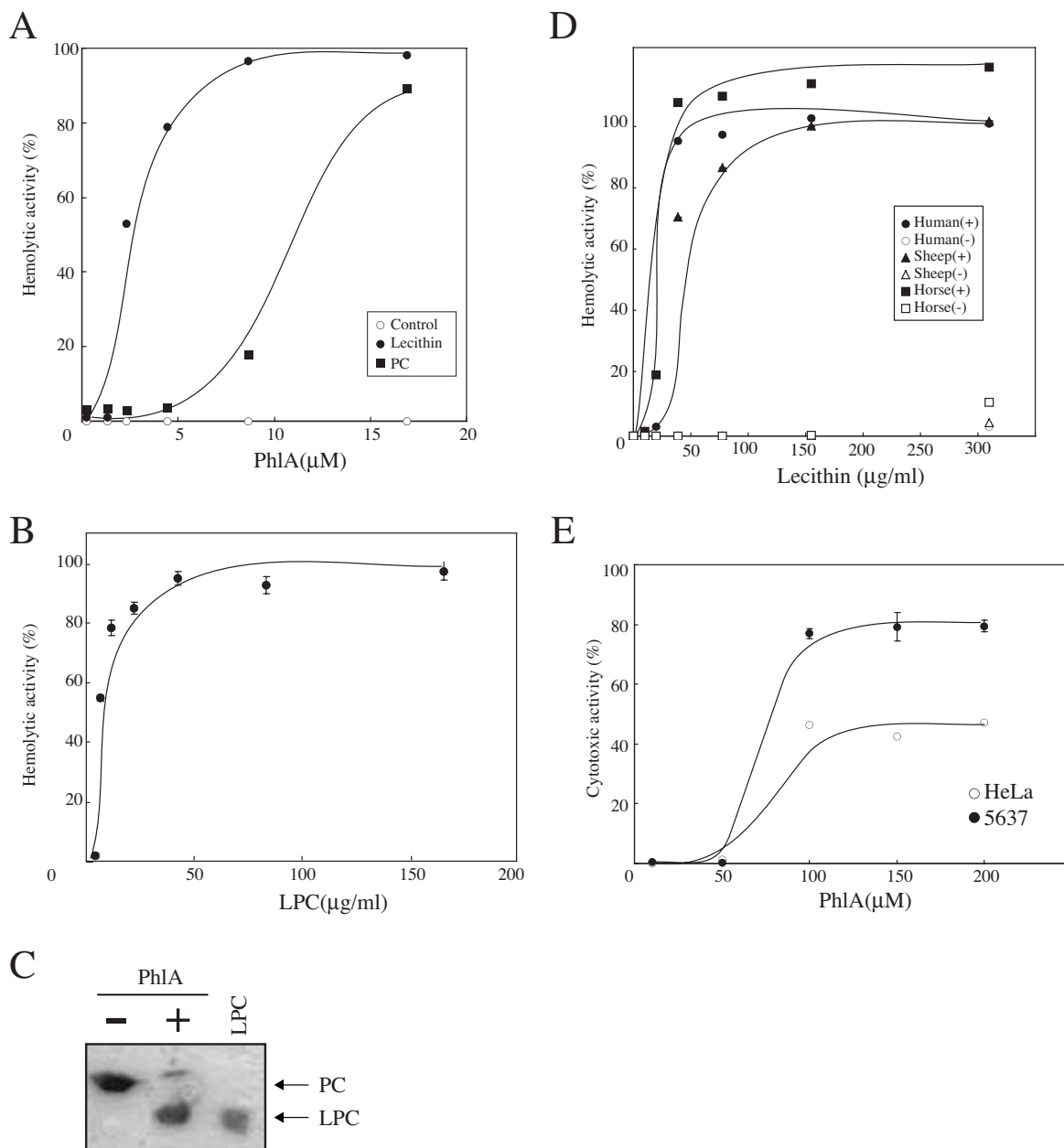


Figure 4
Phospholipid requirements of PhIA hemolytic and cytotoxic activities. (A) Human RBC were mixed with various concentrations of His-PhIA in the absence (open circles) or presence of lecithin (filled circles) or phosphatidylcholine (filled squares) and incubated at 37°C for 1 h. (B) Human RBC were mixed with various concentrations of lysophosphatidylcholine (LPC) and incubated at 37°C for 1 h. (C) Products of the reaction of PC with (+) or without (-) His-PhIA were analyzed by thin-layer chromatography. (D) Human (circles), sheep (triangles), and horse (squares) RBC were mixed with 8.3 μM PhIA (filled symbols) or no PhIA (open symbols) and incubated at 37°C for 1 h in the presence of various concentrations of lecithin with 2 mM CaCl₂. (E) HeLa and 5637 cells were exposed to various concentrations of His-PhIA for 1 h in the presence of lecithin. His-PhIA cytotoxicity was evaluated with a CytoTox 96 Non-Radioactive Cytotoxicity Assay kit (Promega). Open and filled circles show HeLa and 5637 cells, respectively. Values are averages ± SE from three independent experiments. (A), (B), and (D) Results are expressed as percent lysis compared with lysis of RBC in distilled water, as in the contact hemolysis assay (Fig. 1).

Tsubokura et al. [40] reported PL-dependent hemolytic activity in a *Y. enterocolitica* culture filtrate. Schmiel et al. [12] independently identified this hemolysin as a lecithin-dependent phospholipase A (YplA). However, there were no data on whether YplA also had cytotoxic activity in the presence of PL, similar to that reported here for *S. marcescens* PhlA.

PhlA cleaved the ester bond of PL at the *sn*-1 site, and produced fatty acids and LPL from several PLs; e.g., PC, PS, PE, and CL (Fig. 2C). LPL production by PL cleavage might explain why PL addition was required for PhlA hemolytic activity of (Fig. 4A), since LPL may act as a surfactant and induce hemolysis. We detected PhlA hemolytic activity on human blood agar, but not on sheep or horse blood agar (Fig. 1A). However, sheep and horse RBC were lysed with purified PhlA in the presence of PL. This difference may be explained if PLs are released from human RBCs during the preparation of blood agar, and then become substrates for added or secreted PhlA resulting in the production of LPL. In agreement with this possibility, we observed hemolysis around bacterial colonies by addition of egg yolk lecithin to sheep and horse blood agar plates (date not shown).

Our results on the mechanism of PhlA cytotoxic activity allowed us to quantitate cytotoxic activity in a liquid assay. Numerous reports have shown that bacterial phospholipases contribute to pathogenesis by directly hydrolyzing host membrane phospholipids and modulation of the host immune system via the production of lipid second messengers (5, 6, 31). Although PhlA did not produce direct cytotoxicity on cultured cells, the pathogenetic role of indirect cytotoxicity via LPL production should be investigated.

It has been reported that *Pseudomonas aeruginosa* ExoU inhibited neutrophil function in the lungs of infected mice [41] and group A *Streptococcus* (GAS) SlaA contributed to colonization of the upper respiratory tract [37]. Furthermore, a PhlA-like phospholipase, *Y. enterocolitica* YplA, has been shown to play a role in bacterial colonization of the intestinal tract and increasing the pathological changes resulting from the host inflammatory response in the mouse model [12]. The high degree of homology between YplA and PhlA suggests that PhlA may also play a role in *S. marcescens* colonization, since *S. marcescens* is thought to be a commensal in the intestinal tract where PLs are supplied by the host diet. The pathogenic role of PhlA remains to be elaborated.

Conclusions

In this report, we have identified a hemolytic and cytotoxic factor in *S. marcescens* other than the previously reported ShlA. This new factor, PhlA, had phospholipase

A1 activity. It is interesting to note that the hemolytic and cytotoxic activity of PhlA depend on the presence of phospholipids in addition to those in target cell membranes and seems to be mediated by the LPL product of the PhlA enzyme. Our results contribute to understanding the pathogenic role of microbial PLA.

Abbreviations

CL: cardiolipin; EDDA: ethylenediamine-N,N'-diacetic acid; LPC: lysophosphatidylcholine; LPL: lysophospholipid; PC: phosphatidylcholine; PE: phosphatidylethanolamine; PI: phosphatidylinositol; PL: phospholipid; PLA: phospholipase A; RBC: red blood cells; SPM: sphingomyelin.

Authors' contributions

KS carried out most of experimental works, and drafted the manuscript. SI performed the genetic studies. NK improved some of the experimental procedures. YG provided the draft genome sequence information. MO conceived the study and co-wrote the manuscript with HW. All authors have read and approved the final manuscript.

Acknowledgements

This work was supported a grant- in-aid from the Ministry of Health, Labor and Welfare of Japan (H21 Shinkou-Ippan). We thank Drs. J Mitobe, T Kawarai, and M Kuroda for technical advice and discussions.

References

1. Yu VL: **Serratia marcescens: historical perspective and clinical review.** *The New England Journal of Medicine* 1979, **300**:887-893.
2. Hejazi A, Falkiner FR: **Serratia marcescens.** *J Med Microbiol* 1997, **46**:903-912.
3. Hertle R: **The family of Serratia type pore forming toxins.** *Curr Protein Pept Sci* 2005, **4**:313-325.
4. Palmer M: **The family of thiol-activated, cholesterol-binding cytolytins.** *Toxicon* 2001, **39**:1681-1689.
5. Shinoda S, Matsuoka H, Tsuchie T, Miyoshi S, Yamamoto S, Taniguchi H, Mizuguchi Y: **Purification and characterization of a lecithin-dependent haemolysin from Escherichia coli transformed by a Vibrio parahaemolyticus gene.** *J Gen Microbiol* 1991, **137**:2705-2711.
6. Walker DH, Feng HM, Popov VL: **Rickettsial Phospholipase A2 as a pathogenic mechanism in a model of cell injury by typhus and spotted fever group rickettsiae.** *Am J Trop Med Hyg* 2001, **65**:936-942.
7. Hertle R, Schwarz H: **Serratia marcescens internalization and replication in human bladder epithelial cells.** *BMC Infect Dis* 2004, **4**:16.
8. Sakurai J, Nagahama M, Oda M: **Clostridium perfringens alpha-toxin: characterization and mode of action.** *J Biochem* 2004, **136**:569-574.
9. Dorrell N, Martino MC, Stabler RA, Ward SJ, Zhang ZW, McColm AA, Farthing MJG, Wren BW: **Characterization of Helicobacter pylori PldA, a phospholipase with a role in colonization of the gastric mucosa.** *Gastroenterology* 1999, **117**:1098-1104.
10. Flieger A, Neumeister B, Cianciotto N: **Characterization of the gene encoding the major secreted lysophospholipase A of Legionella pneumophila and its role in detoxification of lysophosphatidylcholine.** *Infect Immun* 2002, **70**:6094-6106.
11. Grant KA, Belandia IU, Dekker N, Richardson PT, Park SF: **Molecular characterization of pldA, the structural gene for a phospholipase A from Campylobacter coli, and its contribution to cell-associated hemolysis.** *Infect Immun* 1997, **65**:1172-1180.

12. Schmiel DH, Wagar E, Karamanou L, Weeks D, Miller VL: **Phospholipase A of Yersinia enterocolitica contributes to pathogenesis in a mouse model.** *Infect Immun* 1998, **66**:3941-3951.
13. Givskov M, Olsen L, Molin S: **Cloning and expression in Escherichia coli of the gene for extracellular phospholipase A1 from Serratia liquefaciens.** *J Bacteriol* 1988, **170**:5855-5862.
14. Song JK, Kim MK, Rhee JS: **Cloning and expression of the gene encoding phospholipase A1 from Serratia sp. MK1 in Escherichia coli.** *J Biotechnol* 1999, **72**:103-114.
15. Labbate M, Zhu H, Thung L, Bandara R, Larsen MR, Willcox MD, Givskov M, Rice SA, Kjelleberg S: **Quorum-sensing regulation of adhesion in Serratia marcescens MGI is surface dependent.** *J Bacteriol* 2007, **189**:2702-2711.
16. Goluszko P, Nowacki MR: **Extracellular haemolytic activity of Serratia marcescens.** *FEMS Microbiol Lett* 1989, **61**:207-211.
17. Hertle R, Brutsche S, Groeger W, Hobbie S, Koch W, Konninger U, Braun V: **Specific phosphatidylethanolamine dependence of Serratia marcescens cytotoxin activity.** *Mol Microbiol* 1997, **26**:853-865.
18. Poole K, Braun V: **Influence of growth temperature and lipopolysaccharide on hemolytic activity of Serratia marcescens.** *J Bacteriol* 1988, **170**:5146-5152.
19. Saitoh T, Iyoda S, Yamamoto S, Lu Y, Shimuta K, Ohnishi M, Terajima J, Watanabe H: **Transcription of the ehx enterohemolysin gene is positively regulated by GrlA, a global regulator encoded within the locus of enterocyte effacement in enterohemorrhagic Escherichia coli.** *J Bacteriol* 2008, **190**:4822-4830.
20. Datsenko KA, Wanner BL: **One-step inactivation of chromosomal genes in Escherichia coli K-12 using PCR products.** *Proc Natl Acad Sci USA* 2002, **97**:66400-66405.
21. Iyoda S, Watanabe H: **Positive effects of multiple pch genes on expression of the locus of enterocyte effacement genes and adherence of enterohaemorrhagic Escherichia coli O157: H7 to HEP-2 cells.** *Microbiology* 2004, **150**:2357-2371.
22. Nishihara K, Kanemori M, Yanagi H, Yura T: **Overexpression of trigger factor prevents aggregation of recombinant proteins in Escherichia coli.** *Appl Environ Microbiol* 2000, **66**:884-889.
23. Braun V, Günther H, Neuss B, Tautz C: **Hemolytic activity of Serratia marcescens.** *Arch Microbiol* 1985, **141**:371-376.
24. Gene JA, Gomez M, Gutierrez JM, Cerdas L: **Neutralization of hyaluronidase and indirect hemolytic activities of Costa Rican snake venoms by a polyvalent antivenom.** *Toxicon* 1985, **23**:1015-1018.
25. Al-Abdulla IH, Sidki AM, Landon J: **An indirect haemolytic assay for assessing antivenoms.** *Toxicon* 1991, **29**:1043-1046.
26. Hatic SO II, Picking WL, Young BM, Young GM, Picking WD: **Purification and characterization of two active derivatives of recombinant YpIA, a secreted phospholipase from Yersinia enterocolitica.** *Biochem Biophys Res Commun* 2002, **292**:463-467.
27. Kasurinen J, Vanha-Perttula T: **An enzymatic colorimetric assay of calcium-dependent phospholipases A.** *Anal Biochem* 1987, **164**:96-101.
28. Dittmer JC, Lester RL: **A simple, specific spray for the detection of phospholipids on thin-layer chromatograms.** *J Lipid Res* 1964, **15**:126-127.
29. Behl C, Davis JB, Lesley R, Schubert D: **Hydrogen peroxide mediates amyloid beta protein toxicity.** *Cell* 1994, **77**:817-827.
30. Givskov M, Molin S: **Secretion of Serratia liquefaciens phospholipase from Escherichia coli.** *Mol Microbiol* 1993, **8**:229-42.
31. Sitkiewicz I, Stockbauer KE, Sitkiewicz JM: **Secreted bacterial phospholipase A2 enzymes: better living through phospholipolysis.** *Trends Microbiol* 2007, **15**:63-69.
32. Hendrickson HS, Hendrickson EK, Johnson ID, Farber SA: **Intracellularly quenched BODIPY-labeled phospholipid analogs in phospholipase A(2) and platelet-activating factor acetylhydrolase assays and in vivo fluorescence imaging.** *Anal Biochem* 1999, **276**:27-35.
33. Silverman BA, Weller PF, Shin ML: **Effect of erythrocyte membrane modulation by lysolecithin on complement-mediated lysis.** *J Immunol* 1984, **132**:386-391.
34. Scandella CJ, Kornberg A: **A membrane-bound phospholipase A1 purified from Escherichia coli.** *Biochemistry* 1971, **10**:4447-4456.
35. Istvan TS, Coloe PJ: **Phospholipase A in Gram-negative bacteria and its role in pathogenesis.** *Microbiology* 2006, **152**:1263-1274.
36. Finck-Barbançon V, Goranson J, Zhu L, Sawa T, Wiener-Kronish JP, Fleiszig SM, Wu C, Mende-Mueller L, Frank DW: **ExoU expression by Pseudomonas aeruginosa correlates with acute cytotoxicity and epithelial injury.** *Mol Microbiol* 1997, **25**:547-557.
37. Banks DJ, Beres SB, Musser JM: **The fundamental contribution of phages to GAS evolution, genome diversification and strain emergence.** *Trends Microbiol* 2002, **10**:515-521.
38. Phillips RM, Six DA, Dennis EA, Ghosh P: **In vivo phospholipase activity of the Pseudomonas aeruginosa cytotoxin ExoU and protection of mammalian cells with phospholipase A2 inhibitors.** *J Biol Chem* 2003, **278**:41326-41332.
39. Sitkiewicz I, Nagiec MJ, Sumbly P, Butler SD, Cywes-Bentley C, Musser JM: **Emergence of a bacterial clone with enhanced virulence by acquisition of a phage encoding a secreted phospholipase A2.** *Proc Natl Acad Sci USA* 2006, **103**:16009-16014.
40. Tsubokura M, Otsuki K, Shimohira I, Yamamoto H: **Production of indirect hemolysin by Yersinia enterocolitica and its properties.** *Infect Immun* 1979, **25**:939-942.
41. Diaz MH, Shaver CM, King JD, Musunuri S, Kazzaz JA, Hauser AR: **Pseudomonas aeruginosa induces localized immunosuppression during pneumonia.** *Infect Immun* 2008, **76**:4414-4421.

Publish with **BioMed Central** and every scientist can read your work free of charge

"BioMed Central will be the most significant development for disseminating the results of biomedical research in our lifetime."

Sir Paul Nurse, Cancer Research UK

Your research papers will be:

- available free of charge to the entire biomedical community
- peer reviewed and published immediately upon acceptance
- cited in PubMed and archived on PubMed Central
- yours — you keep the copyright

Submit your manuscript here:
http://www.biomedcentral.com/info/publishing_adv.asp

