

RESEARCH ARTICLE

Open Access

Complex sputum microbial composition in patients with pulmonary tuberculosis

Zelin Cui¹, Yuhua Zhou¹, Hong Li², Yan Zhang¹, Shulin Zhang¹, Shenjie Tang^{2*} and Xiaokui Guo^{1*}

Abstract

Background: An increasing number of studies have implicated the microbiome in certain diseases, especially chronic diseases. In this study, the bacterial communities in the sputum of pulmonary tuberculosis patients were explored. Total DNA was extracted from sputum samples from 31 pulmonary tuberculosis patients and respiratory secretions of 24 healthy participants. The 16S rRNA V3 hyper-variable regions were amplified using bar-coded primers and pyro-sequenced using Roche 454 FLX.

Results: The results showed that the microbiota in the sputum of pulmonary tuberculosis patients were more diverse than those of healthy participants ($p < 0.05$). The sequences were classified into 24 phyla, all of which were found in pulmonary tuberculosis patients and 17 of which were found in healthy participants. Furthermore, many foreign bacteria, such as *Stenotrophomonas*, *Cupriavidus*, *Pseudomonas*, *Thermus*, *Sphingomonas*, *Methylobacterium*, *Diaphorobacter*, *Comamonas*, and *Mobilicoccus*, were unique to pulmonary tuberculosis patients.

Conclusions: This study concluded that the microbial composition of the respiratory tract of pulmonary tuberculosis patients is more complicated than that of healthy participants, and many foreign bacteria were found in the sputum of pulmonary tuberculosis patients. The roles of these foreign bacteria in the onset or development of pulmonary tuberculosis should be considered by clinicians.

Keywords: Pulmonary tuberculosis, Sputum, Microbiota, Diversity

Background

Chronic pulmonary tuberculosis poses a global health emergency. It has been known for many centuries and is mainly caused by the bacillus *Mycobacterium tuberculosis*. Many reports have revealed co-infection with different strains or species of *Mycobacterium* in pulmonary tuberculosis patients. Mixed infection with Beijing and non-Beijing strains of *M. tuberculosis* [1] has been reported to mediate the increased reinfection rate in regions with a high incidence of tuberculosis. Similarly, MAC (*Mycobacterium avium* complex) and *M. tuberculosis* coexist in some patients with combined mycobacterial infections [2]. The systems biology concept of persistent infection is that infectious diseases reflect an equilibrium between the host and the pathogen that is established and maintained by a broad network of interactions. These interactions occur

across scales that range from molecular to cellular, to whole organism and population levels [3].

The development of nucleotide sequencing has helped reveal the importance of microbiota to human health [4]. For example, community and microbial ecology-based pathogenic theories have been introduced to explain the relationship between dental plaque and the host [5]. The urine microbiomes of men with sexually transmitted infection were found to be dominated by fastidious, anaerobic and uncultivable bacteria [6]. Furthermore, the microbiota interact with nutrients and host biology to modulate the risk of obesity and associated disorders, including diabetes, obesity inflammation, liver diseases and bacterial vaginosis (BV) [7-10]. Patients with neonatal necrotising enterocolitis have lower microbiota diversity, which is associated with an increase in the abundance of *Gammaproteobacteria* [11]. Ichinohe *et al* revealed that microbiota can regulate the immune defence against respiratory tract influenza A virus infection [12]. Ehlers and Kaufmann also emphasised the association

* Correspondence: tangsj1106@sina.com; xkguo@shsmu.edu.cn

²Tuberculosis Centre for Diagnosis and Treatment, Shanghai Pulmonary Hospital, Tongji University School of Medicine, Shanghai 200433, China

¹Department of Medical Microbiology and Parasitology, Shanghai Jiao Tong University School of Medicine, 200025, Shanghai, China

between chronic diseases and dysbiosis or a disturbed variability of the gut microbiome [13]. In light of the recent discovery of cystic fibrosis associated lung microbiota, Delhaes and Monchy *et al* discussed the microbial community as a unique pathogenic entity [14]. Huang and Lynch emphasised that microbiota, as a collective entity, may contribute to pathophysiologic processes associated with chronic airway disease [15]. Robinson *et al* also suggested the conservation or restoration of the normal community structure and function of host-associated microbiota should be included in the prevention and treatment of human disease [16]. In summary, microbiota are very important to human health, Understanding the microbial composition in the respiratory tract of pulmonary tuberculosis patients may enhance our awareness of microbiota as a collective entity or even collective pathogenic entity, and the role this entity plays in the onset and development of pulmonary tuberculosis.

In this work, we collected 31 sputum samples from pulmonary tuberculosis patients from Shanghai Pulmonary Hospital, and 24 respiratory secretion samples from healthy participants in Shanghai, China as controls, and investigated the composition of the microbiota in the lower respiratory tract of pulmonary tuberculosis patients.

Results and discussion

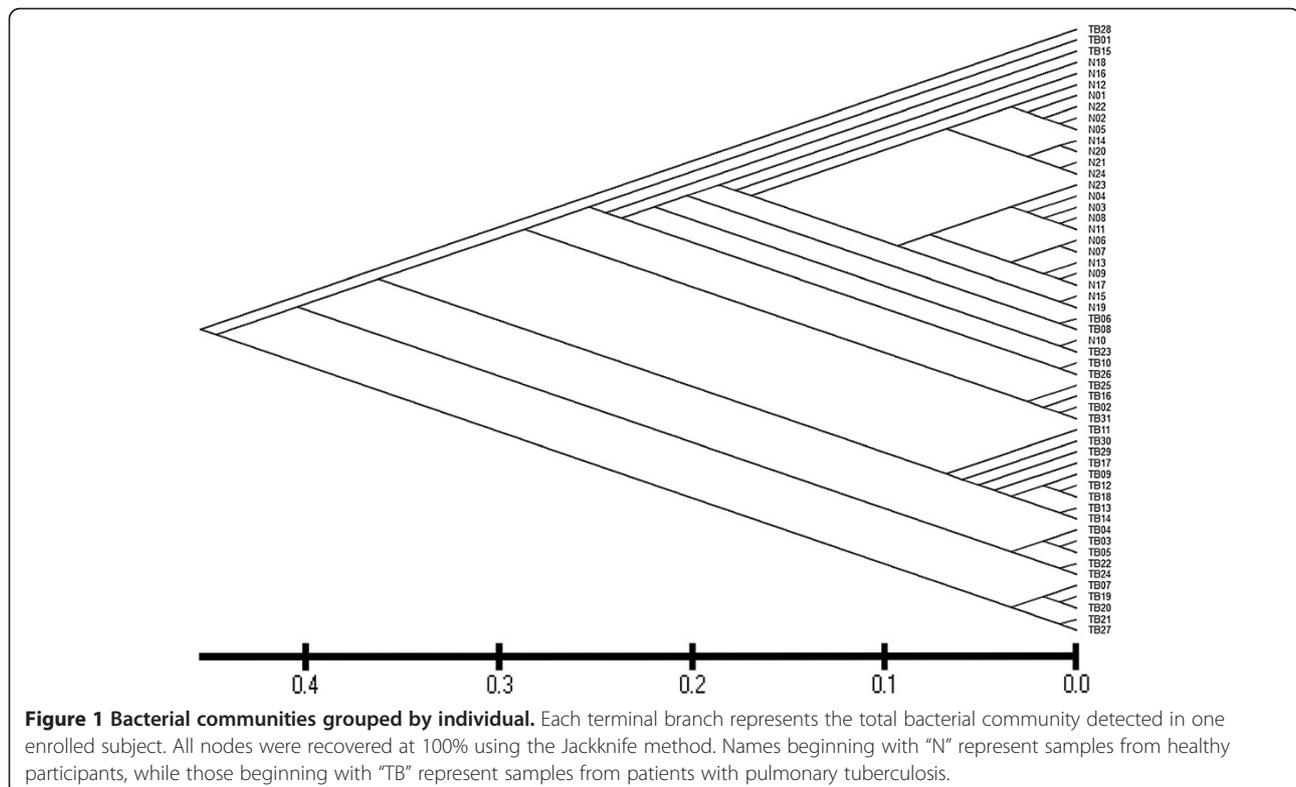
Results

Sequence diversity

The 454 pyro-sequencing method was used to analysis a total of 71928 PCR amplicons in the samples from pulmonary tuberculosis patients and healthy participants, The amplicons averaged approximately 200 bp in length, and the average number of sequences per sample was 1307.8. The mean Shannon diversity and evenness indices in the pulmonary tuberculosis samples were 6.1926 (SD, 0.8093) and 0.9615 (SD, 0.0177), respectively. Both indices were significantly higher than those in the healthy participants, which were 5.5145 (SD, 0.6545) ($p=0.006$) and 0.9341 (SD, 0.0216) ($p=0.000$), respectively.

Clustering analysis of the respiratory tract microbiota can separate healthy participants from pulmonary tuberculosis patients

The similarities between the respiratory tract secretion microbiota of the healthy participants and sputum microbiota of the pulmonary tuberculosis patients were estimated by calculating UniFrac distances. Figure 1 shows that the healthy participants were clustered together, while the pulmonary tuberculosis patients were divided into several different sub-branches.



As shown in Figure 2, clustering after principal coordinate analysis (PCoA) of the UniFrac distance demonstrated a strong clustering of healthy participants away from pulmonary tuberculosis patients. To better characterise the sputum microbiomes, the sequences were sorted to the genera level. A total 614 genera were observed; 235 genera were observed in healthy participants, and 564 genera were found in pulmonary tuberculosis patients, although more than half of these accounted for only a small fraction of the total sequences. As shown in Figure 3, *Streptococcus*, *Granulicatella*, *Actinomyces*, *Prevotella*, and *Veillonella* were predominant in the microbiota of both healthy participants and pulmonary tuberculosis patients. In contrast, *Anoxybacillus*, *Klebsiella*, *Acinetobacter*, *Pilibacter*, *Abiotrophia*, *Paucisalibacillus*, and *Rothia* were more abundant in pulmonary tuberculosis patients than healthy participants. *Neisseria*, *Porphyromonas*, *TM7_genera_incertae_sedis*, *Parvimonas*, *Campylobacter*, *Haemophilus*, and *Fusobacterium* were less common in pulmonary tuberculosis patients than healthy participants. Furthermore, *Stenotrophomonas*, *Cupriavidus*, *Pseudomonas*, *Thermus*, *Sphingomonas*, *Brevundimonas*, *Brevibacillus*, *Methylobacterium*, *Diaphorobacter*, *Comamonas*, *Mobilicoccus*,

and *Fervidicoccus* were unique to and widespread among the pulmonary tuberculosis patients.

The phylum level composition of respiratory microbiomes

A total of 24 phyla were detected in the pulmonary tuberculosis samples, while 17 phyla were detected in healthy participants. Actinobacteria, Bacteroidetes, Proteobacteria, and Crenarchaeota were widely and abundantly distributed among nearly all of the samples. Firmicutes (37.02%), Bacteroidetes (29.01%), Proteobacteria (16.37%), Crenarchaeota (3.16%), and Actinobacteria (2.89%) were common in the healthy participants, while Firmicutes (41.62%), Bacteroidetes (7.64%), Proteobacteria (17.99%), Actinobacteria (21.20%), and Crenarchaeota (7.5%) were common in the pulmonary tuberculosis patients. Chlamydiae, Chloroflexi, Cyanobacteria/Chloroplast, Deinococcus-Thermus, Elusimicrobia, Euryarchaeota, SR1, Spirochaetes, Synergistetes, and Tenericutes were found in both the healthy participants and pulmonary tuberculosis patients, although they were rare in some samples. Aquificae, Caldiseica, Gemmatimonadetes, Lentisphaerae, Planctomycetes, Thermodesulfobacteria, and Verrucomicrobia were unique to the pulmonary tuberculosis samples. Moreover, in healthy

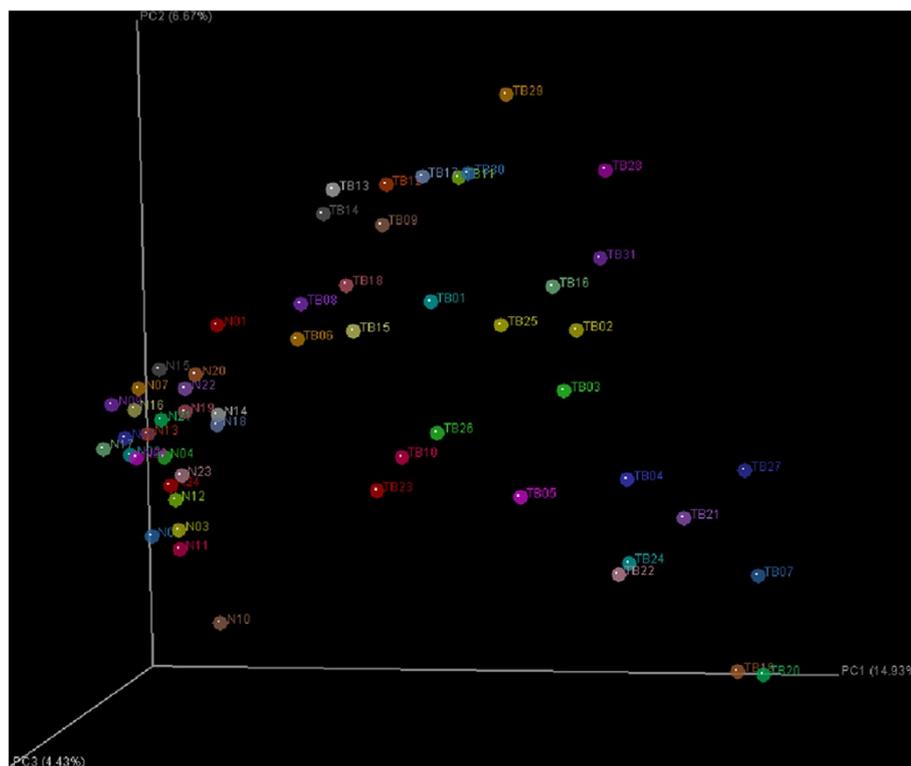


Figure 2 UniFrac community comparison of healthy participants and patients with pulmonary tuberculosis. The sputum microbiomes were clustered using un-weighted UniFrac. The percentage of variation explained by each principal component is indicated on the axis. The circles with names beginning with "N" represent samples from healthy participants, while those beginning with "TB" correspond to samples from patients with pulmonary tuberculosis.

participants, *Deinococcus-Thermus*, Bacteroidetes, and Fusobacteria accounted for 0.01%, 29.01% and 8.06%, respectively. However, in pulmonary tuberculosis patients, *Deinococcus-Thermus* increased to 0.93%, Bacteroidetes, and Fusobacteria decreased to 7.64% and 1.35%, respectively.

Several genera were unique to the respiratory tracts of pulmonary tuberculosis patients

Many genera were unique to in the sputum of pulmonary tuberculosis patients. As shown in Figure 3 and Table 1, *Phenylobacterium*, *Stenotrophomonas*, *Cupriavidus*, and *Pseudomonas* were found in nearly half of the tuberculosis patients we enrolled; furthermore, their total copies accounted for more than 1% of the total sequences from the sputum of pulmonary tuberculosis patients. Other genera such as *Sphingomonas*, *Mobilicoccus*, *Brevundimonas*, *Brevibacillus*, and *Diaphorobacter* were much more widely detected in pulmonary tuberculosis patients, even though they accounted for only a small number of sequences.

Table 1 The distribution of some genera that were uniquely found in the sputum of pulmonary tuberculosis patients

Genera	α	β
<i>Phenylobacterium</i>	13/31	2.15%
<i>Stenotrophomonas</i>	12/31	2.15%
<i>Cupriavidus</i>	16/31	1.60%
<i>Caulobacter</i>	5/31	1.56%
<i>Pseudomonas</i>	15/31	1.27%
<i>Thermus</i>	14/31	0.71%
<i>Sphingomonas</i>	16/31	0.66%
<i>Brevundimonas</i>	17/31	0.49%
<i>Pelomonas</i>	15/31	0.47%
<i>Acidovorax</i>	13/31	0.47%
<i>Brevibacillus</i>	16/31	0.36%
<i>Methylobacterium</i>	13/31	0.34%
<i>Diaphorobacter</i>	17/31	0.31%
<i>Comamonas</i>	14/31	0.26%
<i>Mobilicoccus</i>	20/31	0.24%
<i>Fervidicoccus</i>	13/31	0.21%
<i>Serpens</i>	5/31	0.19%
<i>Lactobacillus</i>	12/31	0.18%
<i>Thermobacillus</i>	12/31	0.16%
<i>Auritidibacter</i>	13/31	0.14%
<i>Deinococcus</i>	9/31	0.13%
<i>Lapillicoccus</i>	13/31	0.11%
<i>Devriesea</i>	13/31	0.11%

" α ": the number of pulmonary tuberculosis patients in whom sequences from the corresponding genera were found.

" β ": the percentage of sequences of the corresponding genera of all sequences found in pulmonary tuberculosis patients.

Several rare genera were present in the sputum of pulmonary tuberculosis patients, such as *Thermus*, *Pelomonas*, *Methylobacterium*, *Comamonas*, *Lactobacillus*, *Thermobacillus*, *Auritidibacter*, *Lapillicoccus*, and *Devriesea*.

Discussion

This study provides the first report on the microbial composition of the lower respiratory tract of pulmonary tuberculosis patients through the amplification of 16S rRNA V3 hyper-variable regions using bar-coded primers and pyro-sequencing by Roche 454 FLX. The results revealed that the microbial composition of the lower respiratory tract in pulmonary tuberculosis patients was more diverse ($p < 0.05$) than in healthy participants. Charlson *et al* reported that the microbial composition of saliva or pharynx secretions can reflect the microbial communities in the lower respiratory tract, and their results showed that there is a topographical continuity of bacterial populations in the healthy human respiratory tract [17]. Therefore, we chose to use sputum and respiratory secretions in this study. However, the best samples to use would be lung lavage fluid, which perfectly reflects the lower microbial composition of the respiratory tract. However, obtaining lung lavage fluid is challenging, especially from healthy volunteers, because lung lavage is painful and may even be harmful. This may raise some ethical issues. In contrast, sputum and respiratory secretions are easily obtained through non-invasive, patients-friendly collection methods. Therefore, we chose to analyse sputum and respiratory tract secretions in our study. A previous study showed that fewer than 1% of commensal organisms are able to grow under laboratory conditions [18]; therefore, traditional cultivation-based strategies for analysing the complexity and genetic diversity of microbial communities are strongly biased. However, modern methods, based on barcoded primers and 454 pyro-sequencing allow for a thorough profiling of the microbiota of each enrolled person [19,20]. Published studies have also proved that the 16S rRNA V3 region sequence ideally suited for distinguishing all bacterial species to the genus level, except for closely related *Enterobacteriaceae* [21].

The lower respiratory tract microbiome of pulmonary tuberculosis patients was distinct from that of the healthy participants. As shown in Figures 1 and 2, the pulmonary tuberculosis patients formed a clear cluster that was separate from the healthy participants based on their microbiota. The phyla Bacteroidetes and Fusobacteria were significantly underrepresented in pulmonary tuberculosis patients compared with healthy participants, while Actinobacteria was significantly overrepresented in pulmonary tuberculosis patients. Moreover, bacteria from the phylum *Deinococcus-Thermus* were widely distributed in pulmonary tuberculosis patients (15/31), but

rarely found in healthy participants, and the phyla Aquificae, Caldiseica, Gemmatimonadetes, Lentisphaerae, Planctomycetes, Thermodesulfobacteria and Verrucomicrobia were unique to pulmonary tuberculosis patients. Figure 1 shows the genera *Klebsiella*, *Pseudomonas* and *Acinetobacter* were more common in pulmonary tuberculosis patients, and we postulated that these bacteria may aggravate the syndrome of pulmonary tuberculosis in these patients. Table 1 shows that the genera *Phenylobacterium*, *Stenotrophomonas*, *Cupriavidus*, *Caulobacter*, *Pseudomonas*, *Thermus* and *Sphingomonas* were unique to and widely distributed in patients with pulmonary tuberculosis.

The respiratory tract microbiota of pulmonary tuberculosis patients, who suffer from chronic infection, might be important in the pathogenicity of this disease. The variety of bacterial genera especially the presence of some abnormal genera in the sputum of pulmonary tuberculosis patients suggested that the pulmonary tuberculosis patient lung is an ecological niche that can support the growth of a high variety of bacteria, especially certain abnormal bacteria. These abnormal genera reportedly widespread in the environment, and some of them have even been reported to be associated with some infectious diseases [22-27]. Coenye et al also reported the isolation of unusual bacteria from the respiratory secretions of cystic fibrosis patients [22]. However, there are few reports on whether these organisms can cause human disease. The lower respiratory tract is an open system and can communicate freely with the environment. We speculated that, in pulmonary tuberculosis patients, the lung micro-environment may become more susceptible to colonisation by some foreign microbes. The host response to pathogens is characterised by rapid recognition combined with strong innate (i.e., inflammatory) and adaptive immune responses, enabling microbial eradication often at the cost of significant tissue damage. Furthermore, the host is constantly facing the challenge of discriminating between symbiotic and pathogenic bacteria to organise an appropriately an adaptive response [28]. These responses lead to the extensive fibrosis associated with recurring infections, possibly leading to a decreased clearance of lymph and lymph-associated particles from the infected region [29]. The lungs of individual patients typically contain diverse lesions with varied overall structures that change over time [3]. Ultimately, a strong host response to the clearance of *M. tuberculosis* may produce local lesions in the lung. This may in turn increase the possibility that foreign bacteria will colonise or grow in the lower respiratory tract. During the initial disease-causing invasion of the lung by *M. tuberculosis*, a strong host immune response may kill or clear some normal bacteria in the lower respiratory tract of pulmonary tuberculosis

patients. This may be why the populations of many normal bacteria are decreased or absent from the microbiota of the pulmonary tuberculosis patients. At the same time, a strong host strong immune response against the pathogen may damage or produce lesions in the lung tissue, and consequently the micro-environment of the lower respiratory tract may favour colonisation or even host invasion by foreign microorganisms. These foreign bacteria may cooperate with *M. tuberculosis* to cause additional damage to the lung tissue. In this model, although *M. tuberculosis* plays a central role in the disease, the other bacteria may assist in the destruction of the lung tissue, especially in active tuberculosis. If *M. tuberculosis* eliminated promptly, however, lung function can be restored. Further investigation will be required to determine whether pulmonary tuberculosis is the cause of increased foreign bacterial colonisation of the lower respiratory tract or vice versa (i.e., the presence of foreign bacteria aggravates the symptoms of pulmonary tuberculosis); is also possible that both occur simultaneously.

Conclusions

This study demonstrated that the microbial composition of the respiratory tract of pulmonary tuberculosis patients was more complicated than that of healthy volunteers, and many foreign bacteria were found in the sputum of pulmonary tuberculosis patients. These foreign bacteria may participate in the onset or development of pulmonary tuberculosis.

Methods

All of the procedures for the collection and handling of patient samples and data were reviewed and approved by the ethics committee of the Shanghai Pulmonary Hospital and Shanghai Jiaotong University School of Medicine, in compliance with the Helsinki Declaration of the World Medical Association. All study subjects provided written informed consent to participate in the study.

Specimens

A total of 31 pulmonary tuberculosis patients, ranging in age 23 to 67 years old, with a age median of 39 years and a male/female ratio of 19/12, were recruited from the Shanghai Pneumonia Hospital. All patients were free of HIV. The patients were clinically diagnosed with pulmonary tuberculosis based on sputum smear, sputum culture, and computed tomography results. The sputum samples were collected after the patients had been admitted to the hospital. A portion of the sputum sample was used for medical tests, and the remaining sputum was preserved for DNA extraction after the patients were confirmed to have pulmonary tuberculosis. Anti-

TB drug treatment (isoniazid, rifampicin, ethambutol, pyrazinamide) and its efficiency as demonstrated by computed tomography, were also considered to confirm the diagnosis of pulmonary tuberculosis. None of the patients had taken antibiotics for at least 3 months before sampling. Of the 31 patients tested, 12 were sputum culture positive, 9 were sputum smear positive, 20 were clinically diagnosed with bilateral tuberculosis, 7 were clinically diagnosed with right pulmonary tuberculosis, 2 were clinically diagnosed with left lung tuberculosis, 1 was clinically diagnosed with tuberculosis pleurisy, and 1 was clinically diagnosed with tuberculosis bronchiectasis.

The healthy volunteers were recruited from the same region as the tuberculosis patients. A total of 24 healthy participants, ranging from 38 to 66 years old, with a median age of 55, and a male and female ratio of 13/11, were recruited from Shanghai, China. The volunteers had similar lifestyle and eating habits, nutritional status and physical condition, were free of basic pulmonary diseases, severe lung disease, severe oral disease, systemic disease and other known diseases such as obesity or diabetes, that could affect the microbial composition of the respiratory tract. Volunteers with a history of smoking or drinking were also excluded. The healthy participants had not taken any antibiotics for at least 3 months before sampling. The samples from healthy participants were a mixture of saliva and pharyngeal secretions collected by deep coughing in the early morning before gargling. By coughing, the community that was originally in the sputum was contaminated by the normal flora of the oral cavity and pharynx. (The detailed information of the pulmonary tuberculosis patients and the healthy participants were showed in Additional file 1).

Establishment of a pyro-sequencing library and pyro-sequencing using the 454 platform

DNA extraction and PCR of the 16S rRNA V3 region were performed as described in our previously published article [20]. However, several additional modifications were made. Fresh sputum samples were chosen soon after routine tests confirmed the diagnosis of pulmonary tuberculosis. After liquefaction at room temperature for 1 hour in a sterilised sodium hydroxide solution, 3 ml of sample was aliquoted into three 1.5 ml Eppendorf tubes, pasteurised at 83°C for 30 min, and further extracted using a Bacterial DNA kit (OMEGA, Bio-Tek, USA). PCR enrichment of the 16S rRNA V3 hyper-variable region was performed with the forward primer 5'-XXXXXXXX-TACGGGAGGCAGCAG-3' and the reverse primer 5'-XX XXXXXX-ATTACCGCGGCTGCTGG-3'. The 5' terminus of each primer contained a different 8-base- oligonucleotide tag (represented by "XXXXXXXX" in the primer

sequence), while the sequence after the hyphen was used to amplify the sequences of the V3 end region.

To ensure that a sufficient quantity of PCR product was amplified, a two-step PCR strategy was used. The first step was carried out in a 25 µl reaction volume containing 2.5 µl of PCR buffer (TAKARA), 0.625 U ExTaq (TAKARA), 0.1 µl of BSA (TAKARA), and 2 µl of primer solution with 100 µmol of each forward and reverse primer and 50 ng of extracted DNA as a template; ddH₂O was added to reach the final volume of the reaction. Touchdown PCR was performed as follows: 5 min at 94°C for initial denaturation, followed by 20 cycles of 1 min at 94°C for denaturation, 1 min at 65°C for annealing and 1 minute at 72°C for extension, with the annealing temperature decreasing by 0.5°C for each cycle. The reaction volume in the second step of the PCR was 50 µl and contained 5 µl of the product from step one as a template. The reaction also included 5 µl of PCR buffer (TAKARA), 1.25 U ExTaq (TAKARA), 0.2 µl of BSA (TAKARA), 24 µl of water and 200 µmol of each barcoded forward and reverse primer. The amplification was carried out for five cycles of 1 minute at 94°C for denaturation, 1 minute at 55°C for annealing, and 1 minute at 72°C, with the temperature maintained at 20°C after the reaction was complete. Sequencing was performed at the Chinese National Human Genome Centre in Shanghai using a Roche 454 FLX instrument. The resulting sequences were published as SRA accession SRA051957.

Phylogenetic and statistical analysis

The datasets were taxonomically grouped using the RDP classifier (the naive Bayesian classifier of the Ribosomal Database Project) at a confidence level of 90% [30]. The gross sequencing data were first searched for the linker, primers, and their reverse complements using the platform provided by the centre. The identified primer sequences were trimmed from each sequence read. Sequence reads that did not contain the 5'-end primer were removed from the dataset. The same program was also used for barcode identification. Barcodes were identified within the first 25 bases of the reads. Sequence reads were binned into FASTA files based on their barcodes.

Individual sequences were aligned using the Aligner tool, and aligned sequences files for each sample were processed by complete linkage clustering using distance criteria. We used Uclust to cluster all of the sequences, with a cut-off value of 97%. After clustering, we used a representative sequence of each type as the OUT (operational taxonomic units), and the record of each OUT sequence included the number of sequences and the associated classification information. These data were used to calculate the Shannon diversity and evenness

indices. Fast UniFrac was used to analyse the phylogenetic microbial communities of the two types of samples [31]. Statistical analyses were carried out in SPSS 19.0, heatmaps were drawn in R, and Shannon diversity indices were estimated using Estimate S Win 8.20.

Additional file

Additional file 1: The detailed information of the pulmonary tuberculosis patients and the healthy participants.

Competing interest

All authors: We do not have any commercial interest in this work and have no conflict of interest with respect to the work represented in this article.

Authors' contributions

ZC analysed the data and wrote and edited the paper; ZC, YZ, and YZ were involved in generating the data; SZ, HL and ST assembled the clinical data and performed sampling; and XG and ST were responsible for the overall concept, design, and conduct of the study. All authors read and approved the final manuscript.

Acknowledgments

We wish to thank the staff of the Chinese National Human Genome Centre in Shanghai. This work was supported in part by the National Natural Science Foundation of China (30770820, 30970125, 81101264 and 811171587), the Program of Shanghai Research and Development (10JC1408200), and the National High Technology Research and Development Program of China Program (2011AA02A119).

Author details

¹Department of Medical Microbiology and Parasitology, Shanghai Jiao Tong University School of Medicine, 200025, Shanghai, China. ²Tuberculosis Centre for Diagnosis and Treatment, Shanghai Pulmonary Hospital, Tongji University School of Medicine, Shanghai 200433, China.

Received: 28 July 2012 Accepted: 5 November 2012

Published: 23 November 2012

References

- Huang HY, Tsai YS, Lee JJ, Chiang MC, Chen YH, Chiang CY, Lin NT, Tsai PJ: Mixed infection with Beijing and non-Beijing strains and drug resistance pattern of *Mycobacterium tuberculosis*. *J Clin Microbiol* 2010, **48**(12):4474–4480.
- Khan Z, Miller A, Bachan M, Donath J: *Mycobacterium Avium* Complex (MAC) Lung Disease in Two Inner City Community Hospitals: Recognition, Prevalence, Co-Infection with *Mycobacterium Tuberculosis* (MTB) and Pulmonary Function (PF) Improvements After Treatment. *Open Respir Med J* 2010, **4**:76–81.
- Young D, Stark J, Kirschner D: Systems biology of persistent infection: tuberculosis as a case study. *Nat Rev Microbiol* 2008, **6**(7):520–528.
- Blaser MJ, Falkow S: What are the consequences of the disappearing human microbiota? *Nat Rev Microbiol* 2009, **7**(12):887–894.
- Kuramitsu HK, He X, Lux R, Anderson MH, Shi W: Interspecies interactions within oral microbial communities. *Microbiol Mol Biol Rev* 2007, **71**(4):653–670.
- Nelson DE, Van Der Pol B, Dong Q, Revanna KV, Fan B, Easwaran S, Sodergren E, Weinstock GM, Diao L, Fortenberry JD: Characteristic male urine microbiomes associate with asymptomatic sexually transmitted infection. *PLoS One* 2010, **5**(11):e14116.
- Delzenne NM, Cani PD: Interaction between obesity and the gut microbiota: relevance in nutrition. *Annu Rev Nutr* 2011, **31**:15–31.
- Wen L, Ley RE, Volchkov PY, Stranges PB, Avanesyan L, Stonebraker AC, Hu C, Wong FS, Szot GL, Bluestone JA, et al: Innate immunity and intestinal microbiota in the development of Type 1 diabetes. *Nature* 2008, **455**(7216):1109–1113.
- Ling Z, Liu X, Chen X, Zhu H, Nelson KE, Xia Y, Li L, Xiang C: Diversity of cervicovaginal microbiota associated with female lower genital tract infections. *Microb Ecol* 2011, **61**(3):704–714.
- Wang Y, Hoenig JD, Malin KJ, Qamar S, Petrof EO, Sun J, Antonopoulos DA, Chang EB, Claud EC: 16S rRNA gene-based analysis of fecal microbiota from preterm infants with and without necrotizing enterocolitis. *ISME J* 2009, **3**(8):944–954.
- Turnbaugh PJ, Ley RE, Mahowald MA, Magrini V, Mardis ER, Gordon JL: An obesity-associated gut microbiome with increased capacity for energy harvest. *Nature* 2006, **444**(7122):1027–1031.
- Ichinohe T, Pang IK, Kumamoto Y, Peaper DR, Ho JH, Murray TS, Iwasaki A: Microbiota regulates immune defense against respiratory tract influenza A virus infection. *Proc Natl Acad Sci U S A* 2011, **108**(13):5354–5359.
- Ehlers S, Kaufmann SH: Infection, inflammation, and chronic diseases: consequences of a modern lifestyle. *Trends Immunol* 2010, **31**(5):184–190.
- Delhaes L, Monchy S, Frealle E, Hubans C, Salleron J, Leroy S, Prevotat A, Wallet F, Wallaert B, Dei-Cas E, et al: The airway microbiota in cystic fibrosis: a complex fungal and bacterial community—implications for therapeutic management. *PLoS One* 2012, **7**(4):e36313.
- Huang YJ, Lynch SV: The emerging relationship between the airway microbiota and chronic respiratory disease: clinical implications. *Expert Rev Respir Med* 2011, **5**(6):809–821.
- Robinson CJ, Bohannon BJ, Young VB: From structure to function: the ecology of host-associated microbial communities. *Microbiol Mol Biol Rev* 2010, **74**(3):453–476.
- Charlson ES, Bittinger K, Haas AR, Fitzgerald AS, Frank I, Yadav A, Bushman FD, Collman RG: Topographical continuity of bacterial populations in the healthy human respiratory tract. *Am J Respir Crit Care Med* 2011, **184**(8):957–963.
- Staley JT, Konopka A: Measurement of in situ activities of nonphotosynthetic microorganisms in aquatic and terrestrial habitats. *Annu Rev Microbiol* 1985, **39**:321–346.
- Han MK, Huang YJ, Lipuma JJ, Boushey HA, Boucher RC, Cookson WO, Curtis JL, Erb-Downward J, Lynch SV, Sethi S, et al: Significance of the microbiome in obstructive lung disease. *Thorax* 2012, **67**(5):456–463.
- Zhou Y, Lin P, Li Q, Han L, Zheng H, Wei Y, Cui Z, Ni Y, Guo X: Analysis of the microbiota of sputum samples from patients with lower respiratory tract infections. *Acta Biochim Biophys Sin (Shanghai)* 2010, **42**(10):754–761.
- Chakravorty S, Helb D, Burday M, Connell N, Alland D: A detailed analysis of 16S ribosomal RNA gene segments for the diagnosis of pathogenic bacteria. *J Microbiol Methods* 2007, **69**(2):330–339.
- Coenye T, Goris J, Spilker T, Vandamme P, LiPuma JJ: Characterization of unusual bacteria isolated from respiratory secretions of cystic fibrosis patients and description of *Inquilinus limosus* gen. nov., sp. nov. *J Clin Microbiol* 2002, **40**(6):2062–2069.
- Chuvochina MS, Marie D, Chevaillier S, Petit JR, Normand P, Alekhina IA, Bulat SA: Community variability of bacteria in alpine snow (Mont Blanc) containing Saharan dust deposition and their snow colonisation potential. *Microbes Environ* 2011, **26**(3):237–247.
- Zhu XH, Li F, Xu JH, Xiang LH, Kang KF: Cutaneous infectious granuloma caused by *Phenylbacterium* in an adult with myelodysplastic syndrome: a first case report. *Am J Clin Dermatol* 2010, **11**(5):363–366.
- Zhang K, Han W, Zhang R, Xu X, Pan Q, Hu X: *Phenylbacterium zucineum* sp. nov., a facultative intracellular bacterium isolated from a human erythroleukemia cell line K562. *Syst Appl Microbiol* 2007, **30**(3):207–212.
- Fishman JA: Infections in immunocompromised hosts and organ transplant recipients: essentials. *Liver Transpl* 2011, **17**(Suppl 3):S34–37.
- Ryan MP, Adley CC: *Sphingomonas paucimobilis*: a persistent Gram-negative nosocomial infectious organism. *J Hosp Infect* 2010, **75**(3):153–157.
- Sansonetti PJ: To be or not to be a pathogen: that is the mucosally relevant question. *Mucosal Immunol* 2011, **4**(1):8–14.
- Ardies CM: Inflammation as cause for scar cancers of the lung. *Integr Cancer Ther* 2003, **2**(3):238–246.
- Cole JR, Wang Q, Cardenas E, Fish J, Chai B, Farris RJ, Kulam-Syed-Mohideen AS, McGarrell DM, Marsh T, Garrity GM, et al: The Ribosomal Database Project: improved alignments and new tools for rRNA analysis. *Nucleic Acids Res* 2009, **37**(Database issue):D141–145.
- Hamady M, Lozupone C, Knight R: Fast UniFrac: facilitating high-throughput phylogenetic analyses of microbial communities including analysis of pyrosequencing and PhyloChip data. *ISME J* 2010, **4**(1):17–27.

doi:10.1186/1471-2180-12-276

Cite this article as: Cui et al.: Complex sputum microbial composition in patients with pulmonary tuberculosis. *BMC Microbiology* 2012 12:276.