

RESEARCH

Open Access



# Optimization of fermentation conditions and medium components for chrysomycin a production by *Streptomyces* sp. 891-B6

Zhe Hu<sup>1,2,3</sup>, Qiangang Weng<sup>1,2,3</sup>, Zhehui Cai<sup>1,2,3</sup> and Huawei Zhang<sup>1,2,3\*</sup>

## Abstract

**Background** Chrysomycin A (CA) is a promising antibiotic for treatment of Gram-positive bacterial infections and cancers. In order to enhance CA yield, optimization of fermentation conditions and medium components was carried out on strain *Streptomyces* sp. 891-B6, an UV-induced mutant with improved CA titer compared with its wide-type marine strain 891.

**Results** Using one-way experiment, the optimal fermentation conditions for CA production in 1-L shake flask were obtained as follows: 12 days of fermentation time, 5 days of seed age, 5% of inoculum volume ratio, 200 mL of loading volume and 6.5 of initial pH. By response surface methodology, the optimal medium components determined as glucose (39.283 g/L), corn starch (20.662 g/L), soybean meal (15.480 g/L) and CaCO<sub>3</sub> (2.000 g/L).

**Conclusion** Validation tests showed that the maximum yield of CA reached 1601.9 ± 56.7 mg/L, which was a 60% increase compared to the initial yield (952.3 ± 53.2 mg/L). These results provided an important basis for scale-up production of CA by strain 891-B6.

**Keywords** Chrysomycin, Marine *Streptomyces*, Fermentation, Optimization, Single-factor experiment, Response surface methodology

## Background

Chrysomycins A-C (CA-CC, Fig. 1) are an unusual class of glycosides with a benzonaphthopyranone structure firstly discovered in 1955 from marine-derived strain *Streptomyces* sp. A-419 [1]. It had been demonstrated CA possesses remarkable antimicrobial activity against

*Mycobacterium tuberculosis* (MT), multi-drug-resistant (MDR) tuberculosis and methicillin-resistant *Staphylococcus aureus* (MRSA) with MIC values of 3.125, 0.4, and 0.05 µg/mL, respectively [2–4], and also displays potent cytotoxic effect on human lymphoblastic leukemia HL-60, KRAS mutation cell NCI-H358 and glioblastoma U251 and U87- MG cell lines with IC<sub>50</sub> values of 0.9, 0.15, 0.475 and 1.77 µM, respectively [5–9]. Therefore, CA has the therapeutic potential for treatment of Gram-positive bacterial infections and cancers.

Strain *Streptomyces* sp. 891 originally from marine sediments had been shown to produce CA-CC with the ratio of 74:22:4 (Fig. 1a) [10]. Although the CA yield of strain 891 had been increased to 3648 ± 119 mg/L using single-factor and orthogonal experiments at 250-mL flask level,

\*Correspondence:

Huawei Zhang  
hwzhang@zjut.edu.cn

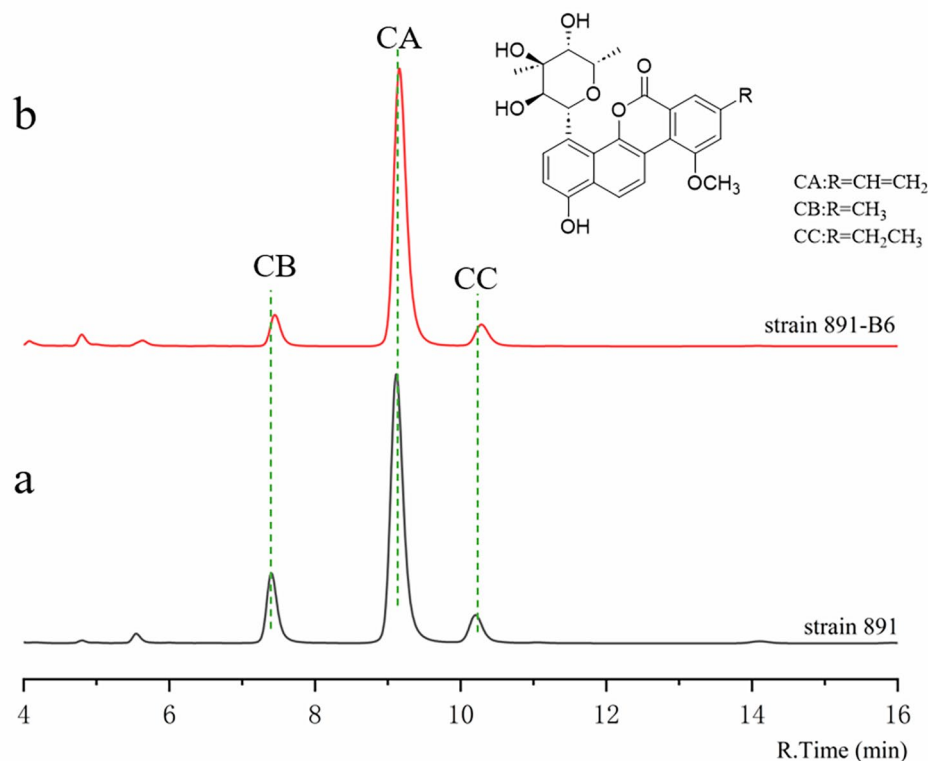
<sup>1</sup>School of Pharmaceutical Sciences, Zhejiang University of Technology, Hangzhou 310014, China

<sup>2</sup>Key Laboratory for Green Pharmaceutical Technologies and Related Equipment of Ministry of Education, Zhejiang University of Technology, Hangzhou 310014, China

<sup>3</sup>Key Laboratory of Pharmaceutical Engineering of Zhejiang Province, Hangzhou 310014, China



© The Author(s) 2024. **Open Access** This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit <http://creativecommons.org/licenses/by/4.0/>. The Creative Commons Public Domain Dedication waiver (<http://creativecommons.org/publicdomain/zero/1.0/>) applies to the data made available in this article, unless otherwise stated in a credit line to the data.



**Fig. 1** Comparison of HPLC profiles of fermentation extract of wide-type strain 891 (a) and UV-mutant strain 891-B6 (b)

the CA content was invariable, causing the high cost of purification process [11]. Strain 891-B6 was obtained as one UV-induced mutant with higher CA content (89%) than that (74%) of the original strain (Fig. 1b) [12], suggesting this mutant is a more ideal strain for producing CA. In order to enhance CA production by strain 891-B6, this work highlighted optimization of fermentation conditions and medium components at 1-L flask level using one-way experiment and response surface methodology.

## Materials and methods

### Strain and medium

Strain 891-B6 was an UV mutant of the wild-type strain of *Streptomyces* sp. 891 and was stored at China General Microbiological Culture Collection Center (CGMCC No.21,775) [13]. ISP-2 was used as basic medium for cultivating strain 891-B6, which consisted of glucose 4 g/L, yeast extract 4 g/L, malt extract 10 g/L and agar 20 g/L.

### Fermentation condition

The seed of strain 891-B6 was prepared using ISP-2 medium and cultivated for 96 h at 30 °C. Before fermentation, an aliquot of 10 mL seed solution was added to each 1-L flask with 200 mL fermentation medium containing corn starch (5.0 g/L), glucose (20 g/L), soybean meal (10 g/L) and CaCO<sub>3</sub> (2.0 g/L). And the initial pH (7.0) was unmodified. The fermentation for CA

production was carried out at 30 °C and 220 rpm in a shaker (ZS-AR, Zhejiang, China) for 10 d.

### Determination of CA yield

By the end of fermentation, 200 mL broth was centrifuged at 4000 rpm (TD5K, Changsha, China) for 20 min and the supernatant was removed. Mycelia of strain 891-B6 was extracted with 800 mL methanol using an ultrasonic extractor (G-080 S, Shenzhen, China) for 20 min at room temperature followed by filtration. An aliquot of 1 mL filtrate was further filtered using organic membrane with diameter of 0.22 μm for HPLC analysis.

### Structure identification of CA

Chemical structure of CA was unambiguously determined by a combination of various spectroscopic methods including H<sup>1</sup>- and C<sup>13</sup>-NMR and ESI-MS as well as comparison with literature data (see supporting material Figs. 1, 2 and 3; Table 1) [5].

### Single-factor experiment

Factors of fermentation condition for CA production were respectively evaluated at various levels, including, seed age from 4 to 9 d, inoculum volume from 2 to 10%, loading volume from 80 to 240 mL, initial pH from 6 to 8.5, glucose concentration from 10 to 50 g/L, corn starch

concentration from 10 to 50 g/L and soybean meal concentration from 5 to 45 g/L.

### Response surface methodology for optimization of medium compositions

Based on the results of the above single-factor experiment, glucose, corn starch and soybean meal concentrations exhibited remarkable effect on CA yield. Therefore, these medium compositions were further optimized for CA production using response surface methodology based on Box-Behnken design (Design Expert 13.0, Stat-Ease Inc., Minneapolis, USA).

### Statistical analysis

Design Expert (version 13.0, Stat-Ease Inc., Minneapolis, USA) was used for analysis of variance (ANOVA) of Box-Behnken design. Each value was expressed as “mean ± standard deviation (SD)”. All experiments were performed three times in parallel.

## Results

### Effect of seed age on CA yield

Seed age is one of important factors affecting product yield since younger or older strain seeds lack strong vitality in their growth and metabolism. When the seed age is short, the formation of mycelial pellets is not conducive to the production of secondary metabolites [14, 15]. The experimental results showed that the highest CA yield (1162.7 ± 75.11 mg/L) achieved at 5-day, and the CA yield decreased significantly owing to mycelial aging (Fig. 2A). Thus, the optimal seed age for CA production is 5-day.

### Effect of inoculum volume on CA Yield

Inoculum amount of strain has an important impact on fermentation process since fewer inoculating volume usually slows down microbial growth and prolongs fermentation time and excessive volume frequently inhibits metabolic level [16]. As shown in Fig. 2B, the CA yield increased in a dependent manner within 2 to 5% of inoculum volume, which the best CA yield was 1035.9 ± 27.34 mg/L.

### Effect of glucose concentration on CA yield

Glucose is one of instant carbon sources for microbial growth and metabolism and its level in broth affects fermentation efficiency [17]. As shown in Fig. 2C, the best CA yield (1139.6 ± 45.6 mg/L) achieved when glucose concentration was 40 g/L. However, it rapidly decreased with the increasing concentration of glucose in fermentation medium.

### Effect of corn starch concentration on CA yield

Starch as macromolecular carbohydrate provides certain nutrients in the later stages of microbial fermentation,

and its appropriate concentration in medium is conducive to biosynthesize secondary metabolites [18]. As shown in Fig. 2D, CA yield reached the highest level (866.3 ± 40.0 mg/L) at corn starch concentration of 20 g/L. However, it gradually declined in a concentration dependent manner within 20 to 50 g/L of starch concentration.

### Effect of soybean meal concentration on CA yield

Soybean meal serves as an important nitrogen source for microbial growth and metabolism. It had been found that excessive nitrogen sources reduce CA yields of the wild strain 891 [19]. As shown in Fig. 3A, the CA yield reached up to 1091.3 ± 63.1 mg/L at the soybean meal concentration of 15 g/L and remarkably decreased as the concentration was higher than 15 g/L.

### Effect of loading volume on CA yield

Various loading volume in a fixed container affects microbial growth and metabolism, and appropriate liquid amount can ensure the demand of oxygen for strains during aerobic fermentation [20]. As shown in Fig. 3B, the highest yield of CA (1162.5 ± 54.18 mg/L) achieved when the loading volume was 200 mL in 1-L flask.

### Effect of initial pH on CA yield

It is well accepted that the optimum pH range for the growth of *Streptomyces* strains is from 6 to 8 [21]. As shown in Fig. 3C, the best CA yield was 1061.3 ± 51.04 mg/L at the initial pH 6.5. But it gradually decreased as the initial pH increased. This is maybe due to the fact that higher pH is unfavourable for those enzymes involved in CA biosynthesis.

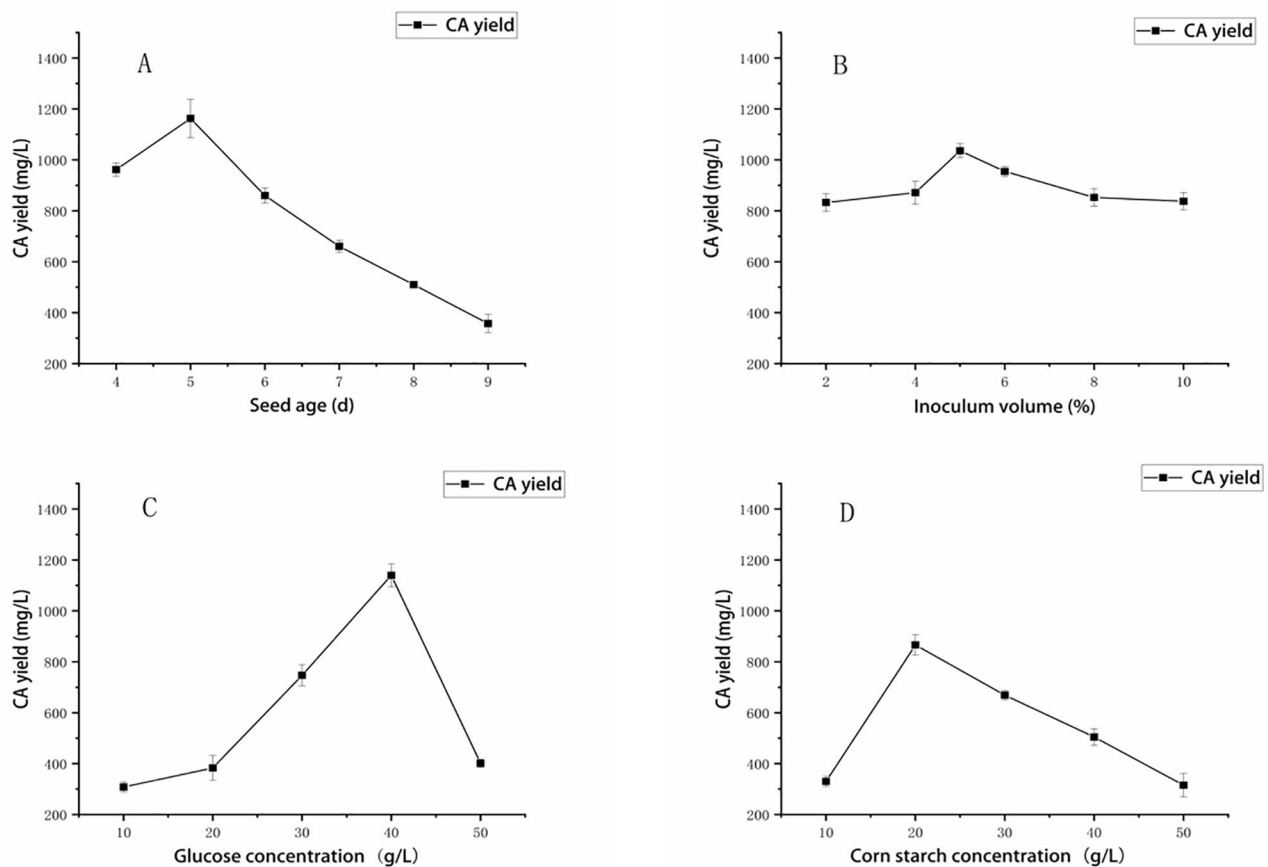
### Response surface methodology

By response surface methodology based on Box-Behnken design, a total of 17 combination experiments were conducted with various concentrations of glucose, corn starch and soybean meal (Table 1). The results were analyzed to afford the following quadratic multinomial regression equation:

$$Y = 1546.40 - 125.96A + 30.49C - 27.95BC - 608.64A^2 - 345.91B^2 - 388.14C^2$$

where Y, A, B and C respectively represent the predicted CA yield, glucose, corn starch, soybean meal.

The ANOVA results shown in Table 2 suggested the regression of the model is highly significant. On basis of the F-value and the magnitude with the P-value, it was concluded that the degree of impact on CA fermentation is in the following order from the largest to the smallest: A > C > B.



**Fig. 2** Effects of fermentation conditions on CA yield of strain 891-B6. (A: seed age; B: inoculum volume; C: glucose concentration; D: corn starch concentration)

The effect of the optimum level of each variable and its interaction on CA yield was investigated by plotting three-dimensional response surfaces and two-dimensional contours for any two independent variables. Under the condition of glucose, corn starch and soybean meal with two certain factors, CA yield gradually increased at the beginning stage and reached the top level at nearby center of each factor, then gradually decreased with the increase of the third factor (Fig. 4). Both contour shapes of factor A (glucose) with factors B (corn starch) and C (soybean meal) were elliptical, indicating significant interactions, while the round contour shapes of factor B with C indicated a moderate interaction (Fig. 5). The model predicted a maximum CA yield of 1552.662 mg/L in 1-L flask when glucose, corn starch and soybean meal were 39.283, 20.662, 15.480 g/L, respectively.

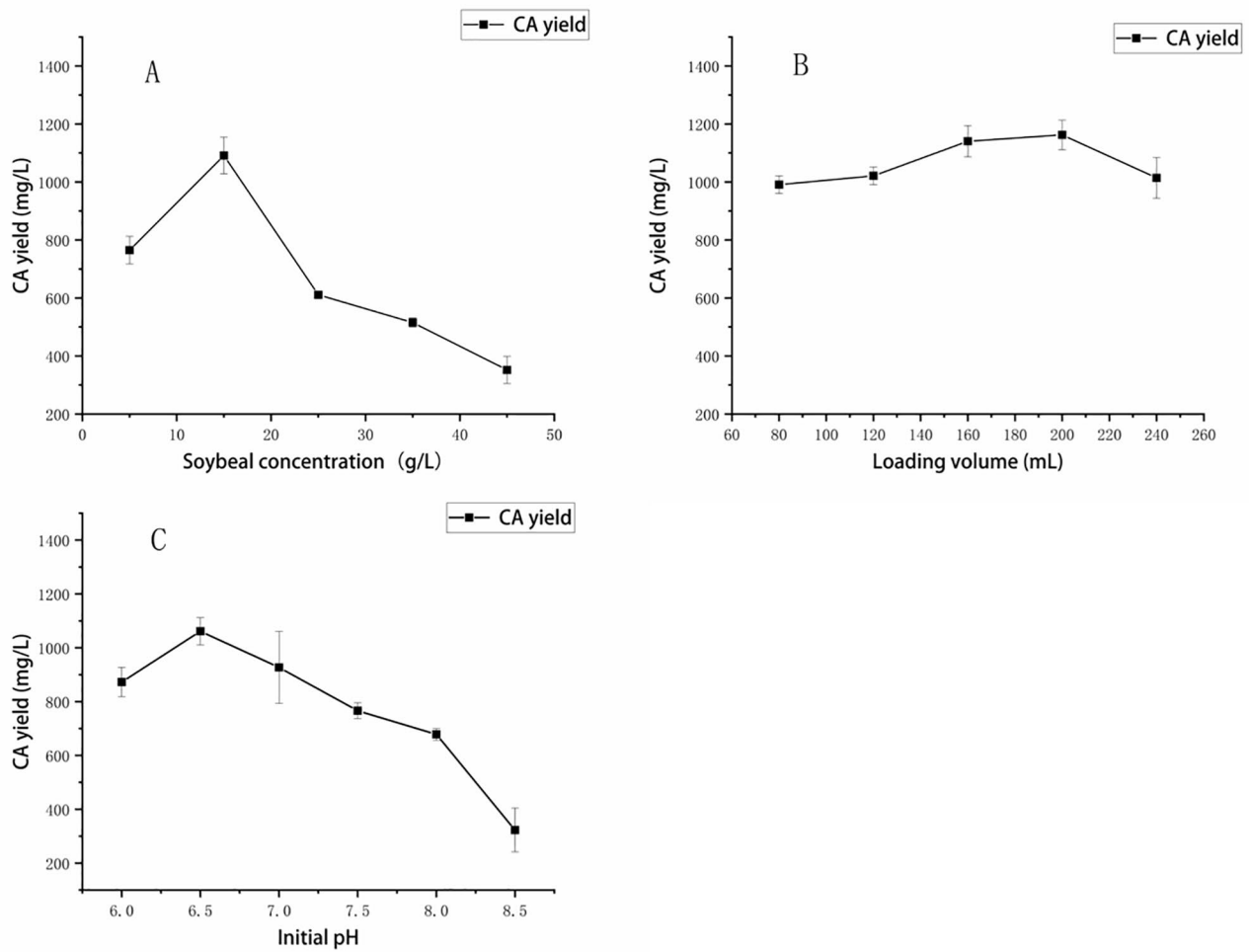
#### Verification result

At a seed age of 5 days, inoculum volume ratio of 5%, loading volume of 200 mL, initial pH 6.5, glucose 39.283 g/L, corn starch 20.662 g/L, soybean meal 15.480 g/L, CaCO<sub>3</sub> 2 g/L, the applicability of the model equations for predicting optimal response values was

tested and fermentation time from 4 to 14 d was examined. As shown in Fig. 6, CA yield gradually increased in a fermentation time-dependent manner and reached the highest level ( $1601 \pm 56.7$  mg/L) at day 12, which was about 60% increase compared with the original titer ( $952.3 \pm 53.2$  mg/L) and showed good agreement with the predicted value (1552.662 mg/L). Therefore, the model developed in this study was adequate for reflecting the predicted optimization of CA production. As of day 12, however, the CA yield began to decrease probably due to the apoptosis of mycelia and/or CA breakdown [22]. So, the best fermentation time for CA production was determined as 12-day.

#### Discussion

Antimicrobial resistance (AMR) has posed a global threat to humankind and could lead to annual deaths up to 10 million people by 2050 [23–26]. Vancomycin is one of the last-line antibacterial agents to treat MRSA infections for nearly four decades, and almost 20 years later several vancomycin-resistant *S. aureus* (VRSA) isolates had been discovered [27, 28]. However, clinical cases of VRSA (with MIC  $\geq 16$   $\mu\text{g/mL}$ ) and vancomycin-intermediate



**Fig. 3** Effects of fermentation conditions on CA yield of strain 891-B6. (A: soybean meal concentration; B: loading volume; C: initial pH)

**Table 1** Experimental design and results of Box-Behnken optimization experiment

Std	Run	Glucose (g/L)	Corn starch (g/L)	Soybean meal (g/L)	CA yield (mg/L)
15	1	40	20	15	1550.5 ± 92.13
9	2	40	10	5	724.7 ± 68.72
16	3	40	20	15	1548.2 ± 83.54
4	4	50	30	15	498.8 ± 42.75
11	5	40	10	25	853.7 ± 88.42
2	6	50	10	15	425.8 ± 34.52
1	7	30	10	15	715.9 ± 48.31
8	8	50	20	25	449.7 ± 22.14
5	9	30	20	5	645.6 ± 56.35
17	10	40	20	15	1549.2 ± 66.46
13	11	40	20	15	1545.2 ± 100.17
6	12	50	20	5	404.8 ± 34.12
7	13	30	20	25	698.4 ± 23.24
10	14	40	30	5	826.9 ± 38.21
12	15	40	30	25	844.1 ± 47.16
3	16	30	30	15	726.9 ± 56.38
14	17	40	20	15	1538.9 ± 34.29

**Table 2** ANOVA for the fitted quadratic polynomial model

Source	Sum of Squares	df	Mean Square	F-value	P-value
Model	3,130,000	9	347,700	4949.17	<0.0001
A-Glucose	126,900	1	126,900	1806.62	<0.0001
B-Corn starch	3898.45	1	3898.45	55.49	0.0001
C-Soybean meal	7435.9	1	7435.9	105.83	<0.0001
AB	961.00	1	961.00	13.68	0.0077
AC	15.6	1	15.6	0.2221	0.6518
BC	3124.81	1	3124.81	44.48	0.0003
A <sup>2</sup>	1,560,000	1	1,560,000	22199.74	<0.0001
B <sup>2</sup>	503,800	1	503,800	7170.72	<0.0001
C <sup>2</sup>	634,300	1	634,300	9028.21	<0.0001
Residual	491.82	7	70.26		
Lack of Fit	406.24	3	135.41	6.33	0.0534
Pure Error	85.58	4	21.39		
Cor Total	3,130,000	16			

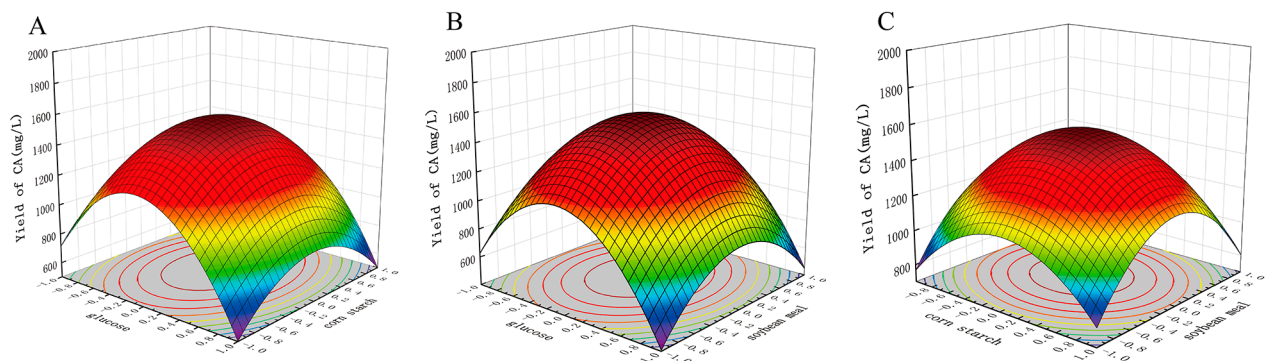
*S. aureus* (VISA) (with MIC > 8 µg/mL) are becoming increasingly common on earth [29]. Therefore, it is urgent to develop novel antibiotics with new actions of mechanism to combat AMR. CA as a drug lead has the

great potential of therapeutic application owing to its potent bactericidal effect on MRSA by targeting multiple critical cellular processes [30]. In this study, optimization of fermentation conditions and medium components for CA production by the modified mutant 891-B6 at flask level were fulfilled by one-way experiments and response surface methodology. These results pave a foundational way for scale-up production of CA and would accelerate the development of new anti-AMR drugs.

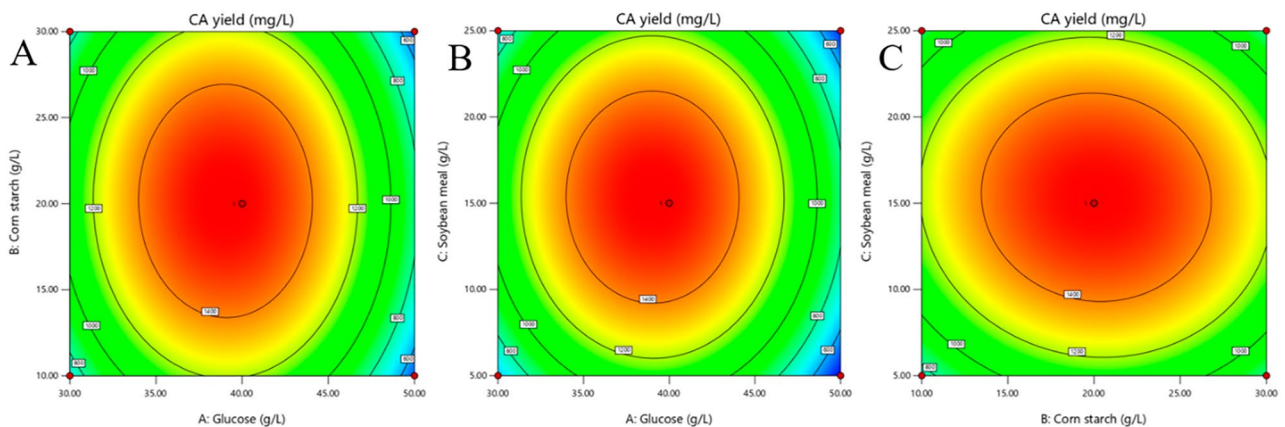
The biosynthesis of CA in several wild strains is usually accompanied by the production of its analogs CB and CC, which pose a great challenge for large-scale production of pure CA. As we know, the increase of target product content effectively reduces its production cost [31, 32]. Therefore, the mutant 891-B6 with higher CA content is more suitable for industrial production of CA.

## Conclusion

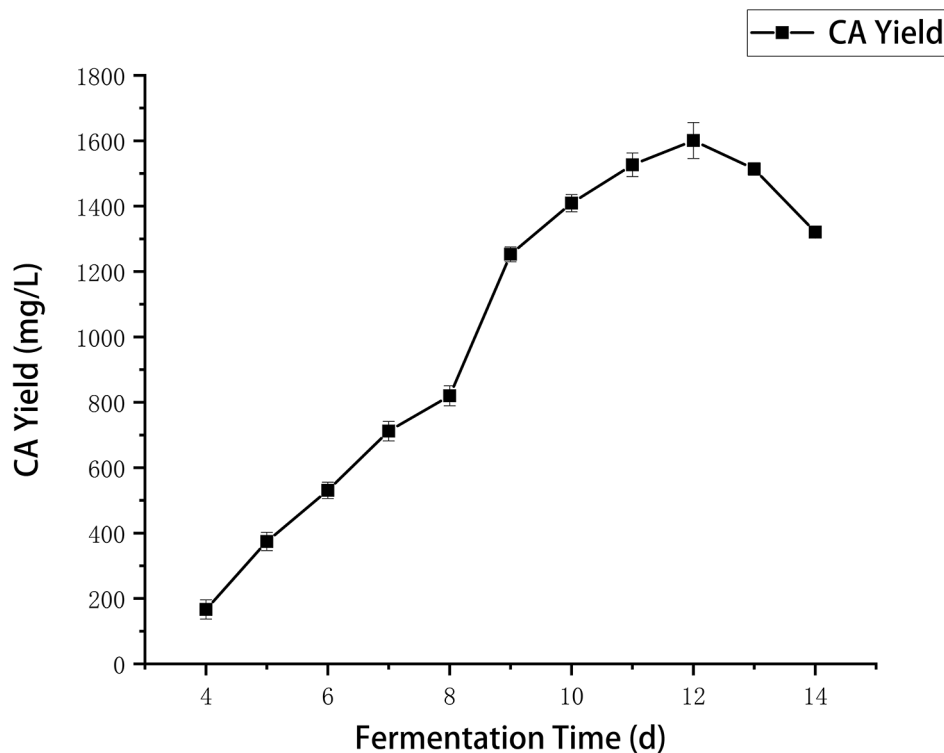
By one-way experiments and response surface methodology, the optimal fermentation conditions and medium formulation for CA production by strain 891-B6 were determined as follows: 5 days of seed age, 5% of inoculum



**Fig. 4** Response surface for CA production by strain 891-B6. (A: interaction between glucose and corn starch; B: interaction between glucose and soybean meal; C: interaction between corn starch and soybean meal.)



**Fig. 5** Two-dimensional contour map for factor interactions. (A: interaction between glucose and corn starch; B: interaction between glucose and soybean meal; C: interaction between corn starch and soybean meal.)



**Fig. 6** Effect of fermentation time on CA yield of strain 891-B6

volume ratio, 200 mL of loading volume, 6.5 of initial pH, 39.283 g/L glucose, 20.662 g/L corn starch, 15.480 g/L soybean meal and 2 g/L  $\text{CaCO}_3$  and 12 days of fermentation time. Under these optimal conditions, the CA yield reached up to  $1601.9 \pm 56.7$  mg/L, which was about 60% increase compared with the original level.

#### Abbreviations

CA-CC	Chrysomycins A-C
MT	<i>Mycobacterium tuberculosis</i>
MDR	Multi-drug-resistant
MRSA	Methicillin-resistant <i>Staphylococcus aureus</i>
NMR	Nuclear Magnetic Resonance Spectroscopy
ESI-MS	Electrospray ionization mass spectrometry
SD	Standard deviation
AMR	Antimicrobial resistance
VRSA	Vancomycin-resistant <i>S. aureus</i>
VISA	Vancomycin-intermediate <i>S. aureus</i>

#### Supplementary Information

The online version contains supplementary material available at <https://doi.org/10.1186/s12866-024-03258-9>.

Supplementary Material 1

#### Acknowledgements

Not applicable.

#### Author contributions

H.W. Zhang contributed to the conceptualization and design of the study. Material preparation, data collection and analysis were performed by Zhe Hu,

Z.H. Cai and Q.G. Weng. The draft of the manuscript was written by Zhe Hu. The final manuscript was read and approved by all authors.

#### Funding

This work was financially supported by the National Key R&D Program of China (2022YFC2804203).

#### Data availability

No datasets were generated or analysed during the current study.

#### Declarations

#### Ethics approval and consent to participate

Not applicable.

#### Consent for publication

Not applicable.

#### Financial interests

The authors have no relevant financial or non-financial interests to disclose.

#### Competing interests

The authors declare no competing interests.

Received: 18 December 2023 / Accepted: 14 March 2024

Published online: 06 April 2024

#### References

1. Strelitz F, Flon H, Asheshov IN. Chrysomycin: a new antibiotic substance for bacterial viruses. *J Bacteriol.* 1955;69:280–3. <https://doi.org/10.1128/JB.69.3.280-283.1955>.
2. Wei TT, Byrne KM, Warnick-Pickle D, Greenstein M. Studies on the mechanism of action of gilvocarcin V and chrysomycin A. *J Antibiot.* 1982;35:545–8. <https://doi.org/10.7164/antibiotics.35.545>.

3. Muralikrishnan B, Dan VM, Vinodh JS, Jamsheena V, Ramachandran R, Thomas S, Dastager S, Kumar KS, Lankalapalli RS, Kumar RA. Anti-microbial activity of chrysomycin A produced by *Streptomyces* sp against *Mycobacterium tuberculosis*. RSC Adv. 2017;7:36335–9. <https://doi.org/10.1039/C7RA05576E>.
4. Wu F, Zhang J, Song FH, Wang SS, Guo H, Wei Q, Dai HQ, Chen XY, Xia XK, Liu XT, Zhang LX, Yu JQ, Lei XG. Chrysomycin A derivatives for the treatment of multi-drug-resistant *tuberculosis*. ACS Cent Sci. 2020;6:928–38. <https://doi.org/10.1021/acscentsci.0c00122>.
5. Weiss U, Yoshihira K, Highet RJ, White RJ, Wei TT. The chemistry of the antibiotics chrysomycin A and B antitumor activity of chrysomycin A. J Antibiot. 1982;35:1194–201. <https://doi.org/10.7164/antibiotics.35.1194>.
6. Jain SK, Pathania AS, Parshad R, Raina C, Ali A, Gupta AP, Kushwaha M, Aravinda S, Bhushan S, Bharate SB, Vishwakarma RA. Chrysomycins A-C, antileukemic naphthocoumarins from *Streptomyces sporoverrucosus*. RSC Adv. 2013;3:21046–53. <https://doi.org/10.1039/C3RA42884B>.
7. Wada S-i, Sawa R, Iwanami F, Nagayoshi M, Kubota Y, Iijima K, Hayashi C, Shibuya Y, Hatano M, Igarashi M, Kawada M. Structures and biological activities of novel 4'-acetylated analogs of chrysomycins A and B. J Antibiot. 2017;70:1078–82. <https://doi.org/10.1038/ja.2017.99>.
8. Liu DN, Liu M, Zhang SS, Shang YF, Song FH, Zhang HW, Du GH, Wang YH. Chrysomycin A inhibits the proliferation, migration and invasion of U251 and U87-MG glioblastoma cells to exert its anti-cancer effects. Molecules. 2022;27:6148. <https://doi.org/10.3390/molecules27196148>.
9. Zhang JM, Liu P, Chen JW, Yao DH, Liu Q, Zhang JH, Zhang HW, Leung ELH, Yao XJ, Liu L. Upgrade of chrysomycin A as a novel topoisomerase II inhibitor to curb KRAS-mutant lung adenocarcinoma progression. Pharmacol Res. 2023;187:106565. <https://doi.org/10.1016/j.phrs.2022.106565>.
10. Hu X, Tang YQ, Liu YY, Pei XY, Huang ZW, Song FH, Zhang HW. Comprehensive genomic analysis of marine strain *Streptomyces* sp. 891, an excellent producer of chrysomycin A with therapeutic potential. Mar Drugs. 2022;20:287. <https://doi.org/10.3390/md20050287>.
11. Ni HJ, Lv SY, Sheng YT, Wang H, Chu XH, Zhang HW. Optimization of fermentation conditions and medium compositions for the production of chrysomycin A by a marine-derived strain *Streptomyces* sp. 891. Prep Biochem Biotechnol. 2021;51:998–1003. <https://doi.org/10.1080/10826068.2021.1885046>.
12. Zhang H, Lv S, Tang Y, Zhu W, Wang H. Streptomycete Mutant and Application. CN Patent 113278545 A, 20 August 2021.
13. Zhu WJ, Pei XY, Chen XY, Wu Y, Song FH, Zhang HW. Comparative transcriptome analysis of two chrysomycin-producing wild-type and mutant strains of *Streptomyces* sp. 891. Metabolites. 2022;12:1170. <https://doi.org/10.3390/metabo12121170>.
14. Yan YZ, Wei XD, Na K, Jin YY, Zhao B, Zhao WJ. Mutation breeding and optimization of fermentation for high production of natamycin by *Streptomyces Natalensis*. Zhongguo Kangshengsu Zazhi. 2013;38:332–8.
15. Kumar P, Khushboo, Rajput D, Dubey KK. Insights into the mechanism of mycelium transformation of *Streptomyces toxytricini* into pellet. FEMS Microbiol Ecol. 2023;4:xtad017. <https://doi.org/10.1093/femsmc/xtad017>.
16. Fu XQ, Wei SJ, Huang WX, Tu GQ. A study on conditions for fermentation of *Streptomyces nanchangensis*. Acta Agric Universitatis Jiangxiensis. 2007;29:457–60.
17. Chang YH, Chang KS, Chen CY, Hsu CL, Chang TC, Hung-Der J. Enhancement of the efficiency of bioethanol production by *saccharomyces cerevisiae* via gradually batch-wise and fed-batch increasing the glucose concentration. Fermentation. 2018;4:45. <https://doi.org/10.3390/fermentation4020045>.
18. Liu GZ, Shen JQ. Effects of culture and medium conditions on hydrogen production from starch using anaerobic bacteria. J Bioscience Bioeng. 2004;98:251–6. [https://doi.org/10.1016/S1389-1723\(04\)00277-4](https://doi.org/10.1016/S1389-1723(04)00277-4).
19. Jiang LF. Effect of nitrogen source on curdlan production by *Alcaligenes Faecalis* atcc 31749. International. J Biol Macromol. 2013;52:218–210. <https://doi.org/10.1016/j.jbiomac.2012.10.010>.
20. Xiong ZQ, Tu XR, Tu GQ. Studies on unsterile fermentation process of hong-gumycin. J Chin Biotechnol. 2008;28:76–9.
21. Fatokun EN, Nwodo UU, Okoh AI. Classical optimization of cellulase and xylanase production by a marine *Streptomyces* Species. Appl Sciences-Basel. 2016;6:286. <https://doi.org/10.3390/app6100286>.
22. Fan XY, Cheng QM, Chen YL, Li P, Long JH, Li MY, Chen C. Effects of different storage methods and fermentation time on conventional nutrient contents, fermentation quality and microbial diversity of moutai distillers grains. Chin J Anim Nutr. 2022;34:5283–94.
23. Jubeh B, Breijyeh Z, Karaman R. Antibacterial prodrugs to overcome bacterial resistance. Molecules. 2020;25:1543. <https://doi.org/10.3390/molecules25071543>.
24. O'Neill J. Tackling drug-resistant infections globally: final report and recommendations. London: Review on Antimicrobial Resistance. 2016. <https://wellcomecollection.org/works/thwvsuba>.
25. O'Neill J. Antimicrobial resistance: tackling a crisis for the health and wealth of nations. London: Review on Antimicrobial Resistance. 2014. <https://wellcomecollection.org/works/rdpck35v>.
26. Murray CJL, Ikuta KS, Sharara F, Swetschinski L, Aguilar GR, Gray A. Global burden of bacterial antimicrobial resistance in 2019: a systematic analysis. Lancet. 2022;10325:399. [https://doi.org/10.1016/S0140-6736\(21\)02724-0](https://doi.org/10.1016/S0140-6736(21)02724-0).
27. Werner GSB, Witte W. Acquired Vancomycin resistance in clinically relevant pathogens. Future Microbiol. 2008;3:547–62. <https://doi.org/10.2217/17460913.3.5.547>.
28. Shariati A, Dadashi M, Taati M, Sarokhalil DD. Global prevalence and distribution of Vancomycin resistant, Vancomycin intermediate and heterogeneously Vancomycin intermediate *staphylococcus aureus* clinical isolates: a systematic review and meta-analysis. New Microbes New Infections. 2020;10:12689. <https://doi.org/10.1038/s41598-020-69058-z>.
29. Chambers HF, Deleo FR. Waves of resistance: *Staphylococcus aureus* in the antibiotic era. Nat Rev Microbiol. 2009;7:629–41. <https://doi.org/10.1038/nrmicro2200>.
30. Jia J, Zheng MX, Zhang CW, Li BL, Lu C, Bai YF, Tong Q, Hang XD, Ge YX, Zeng LP, Zhao M, Song FH, Zhang HW, Zhang L, Hong K, Bi HK. Killing of *Staphylococcus aureus* persists by a multitarget natural product chrysomycin A. Sci Adv. 2023;9:31. <https://doi.org/10.1126/sciadv.adg5995>.
31. Bagherzadeh A, Smejkal Q, Freißlich U, Pfauntsch J, Gruden S, Howe S. Process optimization and reduction of production costs in modern beet sugar manufacturing. Sugar Ind. 2023;148:11: 691–9. <https://doi.org/10.36961/si30577>.
32. Braga A, Faria N. Bioprocess optimization for the production of aromatic compounds with metabolically engineered hosts: recent developments and future challenges. Front Bioeng Biotechnol. 2020;8. <https://doi.org/10.3389/fbioe.2020.00096>.

## Publisher's Note

Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.